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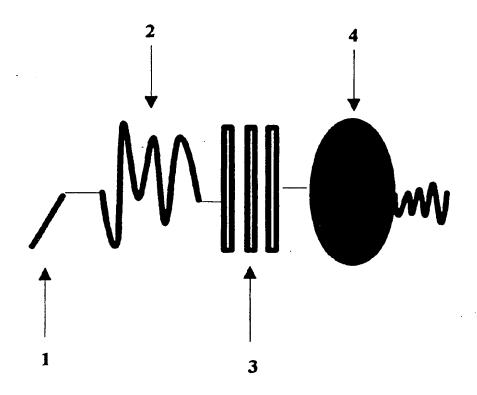
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(54) Title: HYBRID PROTEINS HAVING CROSS-LINKING AND TISSUE-BINDING ACTIVITIES

(57) Abstract

Hybrid proteins having crosslinking and tissue-binding activities, DNA molecules encoding such proteins and methods for producing the hybrid proteins from recombinant host cells are disclosed. The hybrid proteins disclosed herein are useful in tissue sealant and wound healing formu-



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Description

Hybrid Proteins Having Cross-Linking and Tissue-Binding Activities

5 <u>Technical</u> Field

The present invention relates generally toward methods for producing recombinant hybrid proteins, and more specifically, to methods for producing hybrid proteins from host cells through the use of recombinant DNA techniques.

Background of the Invention

The utilization of tissue sealants to replace or augment the use of mechanical wound closure devices has expanded in recent years in many surgical and trauma applications. Tissue sealants include biological adhesives (e.g. fibrin-based adhesives) and synthetic preparations (e.g. cyanoacrylates). Ιt is acknowledged that the use of synthetic preparations of tissue sealants is limited due to their toxicity and limited applications. Biological tissue adhesives have demonstrated utility in cases where the use of mechanical devices to close wounds is insufficient, such joining blood vessels, closing holes in the dura, and in surgery on small or delicate tissues such as in the eye or ear.

Fibrin-based biological tissue generally contain fibrinogen, factor XIII and thrombin as principal ingredients, although in practice biological tissue adhesives are derived from whole blood and contain additional blood proteins. The fibrinogen and factor XIII components of these adhesives are prepared from pooled human plasma by cryoprecipitation (e.g. U.S. Patents No. 4,377,572; 4,362,567; 4,909,251), by ethanol precipitation (e.g. U.S. Patent No. 4,442,655) or from single donor plasma (e.g. U.S. Patent No. 4,627,879; Spotnitz et al., Am. Surq. <u>55</u>: 166-168, 1989). The resultant

fibrinogen/factor XIII preparation is mixed with bovine thrombin immediately before use to convert the fibrinogen to fibrin and activate the factor XIII, thus initiating coagulation of the adhesive.

Fibrin-based tissue adhesives, in their current form, have significant drawbacks that include standardization, lack of quality control from batch the possibility of transmission of human immunodeficiency virus (HIV), hepatitis virus and other etiologic agents. While recombinant production thrombin and factor XIII have been reported, and while proteins might be used in biological tissue adhesives, the biological tissue adhesives still rely on large amounts of fibrinogen that is obtained from pooled human blood. At present, current fibrin(ogen)-based tissue adhesives are not approved for use in the United States.

There is therefore a need in the art for tissue adhesive components, particularly components that facilitate cross-linking to improve clot strength, that are prepared at high levels with reproducible activity and which do not carry the possibility transmission of viral or other etiologic agents. invention addresses these needs by providing recombinant hybrid proteins that provide cross-linking and tissue-adhesive properties and that may be prepared at high levels.

Disclosure of the Invention

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Briefly stated, the present invention provides hybrid proteins having cross-linking and tissue-binding activities, DNA molecules encoding such hybrid proteins and methods for producing hybrid proteins by recombinant means. In one aspect, In one aspect of the invention, the hybrid proteins comprise a tissue-binding domain from a first protein covalently linked to a cross-linking domain from a second protein. Within a related aspect of the

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invention, the tissue-binding domain of the first protein is a heparin binding domain of thrombospondin, a heparin binding domain of fibronectin, a collagen binding domain of fibronectin or a cell binding domain of fibronectin. Within a preferred embodiment, the tissue-binding domain of the first protein comprises the amino acid sequence of Sequence ID No. 6 from Alanine, amino acid 2 to Glutamic acid, amino acid number 926. Within another related aspect of the invention, the cross-linking domain of the second protein comprises the carboxy-terminal 103 amino acids of loricrin, the ten amino acid repeat beginning with glutamine amino acid number 496 of involucrin or the 400 amino-terminal amino acids of the fibrinogen α chain. a preferred embodiment of the invention, tissue-binding domain of the second protein comprises the amino acid sequence of Sequence ID No. 6 from Glycine, amino acid number 928 to Proline, amino acid number 1336. Within a particularly preferred embodiment, the hybrid protein comprises the amino acid sequence of Sequence ID No. 6 from alanine, amino acid number 2 to proline, amino acid number 1336.

The present invention provides DNA molecules encoding hybrid proteins of the present comprising a first DNA segment encoding a tissue-binding domain from a first protein joined to a second DNA segment encoding a cross-linking domain from a second protein. embodiment, the first DNA segment comprises nucleotide sequence of Sequence ID No. 5 from nucleotide 3 to nucleotide 2780. In another embodiment, the second DNA segment comprises the nucleotide sequence of Sequence ID No. 5 from nucleotide 2784 to nucleotide 4013. preferred embodiment, the DNA molecule comprises the nucleotide sequence of Sequence ID Number from nucleotide 3 to nucleotide 4013.

In related embodiments of the invention, DNA constructs are provided which comprise a DNA molecule encoding a hybrid protein, whereins said DNA molecule

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comprises a first DNA segment encoding a tissue-binding domain from a first protein joined to a second DNA segment encoding a cross-linking domain from a second protein and wherein said DNA molecule is operably linked to other DNA segments required for the expression of the DNA molecule. Other embodiments of the invention concern host cells containing the DNA constructs of the present invention and methods of producing hybrid proteins.

10 <u>Brief Description of the Drawings</u>

Figure 1 discloses a representative hybrid protein containing (1) an N-terminal end-to-end interchain cross-linking domain, (2) a domain that promotes inter-chain cross-linking; (3) a domain that confers tissue binding activity; and (4) a carboxy-terminal domain that promotes end-to-end inter-chain cross-linking.

Figures 2-5 disclose absorbance time courses of representative cross-linking assays carried out in the presence of varying levels of factor XIII (activated to factor XIIIa via thrombin during the assay) or factor XIIIa.

Detailed Description of the Invention

present invention provides novel The proteins having cross-linking and tissue activities. The hybrid proteins comprise a cross-linking domain from a first protein covalently linked to a tissuebinding domain from a second protein. The hybrid proteins of the present invention are capable of cross-linking to themselves and to other proteins such as fibrin fibrinogen and are capable of adhering to cell surfaces and/or extracellular matrix components. While not wishing to be bound by a graphical representation, Figure 1 shows a representative hybrid protein containing an N-terminal end-to-end inter-chain cross-linking domain; a domain that promotes inter-chain cross-linking; a domain that confers tissue binding activity; and a carboxy-terminal domain

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that promotes end-to-end inter-chain cross-linking. As used herein, cross-linking refers to the formation of covalent bonds between polypeptides.

The hybrid proteins of the present invention are useful as components of tissue sealant formulations to provide matrix material and to improve clot strength over a wound site, and as components in formulations that promote wound healing. The proteins of the present invention may contain native (i.e. wild-type) protein domains as well as domains that are allelic variants and genetically engineered or synthetic variants respective naturally occurring domains. Such variants are characterized by the presence of conservative amino acid substitutions and/or other minor additions, substitutions or deletions of amino acids.

used within the context As of the invention, tissue-binding domains include protein domains containing amino acid sequences that facilitate adherence to cell surfaces and/or to extracellular matrix components such as collagen, fibronectin, hyaluronic acid glycosaminoglycans. Fibronectin, for example, contains the sequence Gly-Arg-Gly-Asp-Ser (from amino acid 1614 through amino acid 1618 of Sequence I.D. No. 3) that has been shown to be central to cell recognition fibronectin receptor (for review see Yamada, Opinion in Cell Biology 1: 956-963, 1989). The heparin binding domains of fibronectin (Sekiguchi et al., Proc. Natl. Acad. Sci. USA 2661-2665, <u>77</u>: 1980), thrombospondin (Zardi et al., EMBO J. 6: 2337-3342, 1987 and Gutman and Kornblihtt, Proc. Natl. Acad. Sci. USA 84: 7179-7182, 1987) contain sequences that recognize heparin sulfate-containing glycosaminoglycans which extracellular matrix components. The collagen binding domain of fibronectin (Sekiguchi et al. ibid., 1980) contains amino acid sequences that bind to the extracellular matrix component collagen.

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Particularly preferred tissue-binding are the heparin binding domain of fibronectin, comprising the sequence of amino acids of Sequence I.D. No. 2 from alanine, amino acid number 1812 to valine, amino number 2171; the collagen binding domain of fibronectin, comprising the sequence of amino acids of Sequence I.D. No. 2 from glycine, amino acid number 282 to serine, amino acid number 608; and the amino terminal 229 amino acids of thrombospondin. In this regard, a particularly preferred tissue-binding domain is the cell-binding fibronectin, comprising the sequence of amino acids of Sequence I.D. No. 3 from alanine, amino acid number 1357 to glutamic acid, amino acid number 1903. It will be evident to one skilled in the art that smaller portions of the cell-binding domain of fibronectin may be used within hybrid proteins of the present invention, particularly the sequence of amino acids of Sequence I.D. 3 from isoleucine, number 1532 through threonine, amino acid number 1631. As noted above, it is generally accepted that the sequence Gly-Arg-Gly-Asp-Ser acids 1614 to 1618 of Sequence I.D. No. 3) is central to cell recognition by fibronectin.

Cross-linking domains suitable for use in the hybrid proteins of the present invention are protein domains which contain amino acid sequences required for the formation of specific covalent bonds between peptide chains. In a preferred embodiment the inter-chain crosslinks are covalent bonds formed by the action transglutaminase such as XIII, factor tissue transglutaminase, prostate transglutaminase, keratinocyte transglutaminase, epidermal transglutaminase or placental transglutaminase. Transglutaminases catalyze formation of ϵ -(γ -glutamyl)lysine bonds between specific glutamine and lysine residues. However, other inter-chain cross-links, such as those formed by disulfide bonds, are also suitable cross-links. Suitable cross-linking domains include domains from the fibrinogen α chain,

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glutamine/lysine rich domains of loricrin that involved in isodipeptide cross-link formation (Hohl al., <u>J. Biol. Chem.</u> <u>266</u>: 6626-6636, 1991), and at least one of the 10 amino acid-long repeats of involucrin (Cell 5 46: 583-589, 1986 and Etoh et al., Biochem. Biophys. Res. Comm. 136: 51-56, 1986). Preferred cross-linking domains are the carboxy-terminal 103 amino acids of loricrin (Hohl et al., ibid.) and the ten-amino acid repeat beginning with glutamine, amino acid number 496 of involucrin (Simon 10 (J. Biol. Chem. 263: 18093-18098, particularly preferred cross-linking domain comprises the 400 amino-terminal amino acids of the fibrinogen α chain (Doolittle et al., <u>Nature</u> 280: 464-468, 1979; Rixon et **Biochemistry** 22: 3250-3256, 1983). 15 particularly, the amino acid sequence of Sequence ID No. 6 from Glycine, amino acid number 928 to Proline, amino acid number 1336 is preferred.

Although the hybrid proteins of the present invention may consist essentially of covalently linked 20 cross-linking and tissue binding domains, they may further contain domains that facilitate end-to-end covalent cross-The γ chain of fibrinogen contains a domain that facilitates end-to-end cross-linking to another γ chain via ϵ -(γ -glutamyl)lysine bonds. This domain includes at least the 19 carboxy-terminal amino acids and more preferably includes the amino-terminal 275 amino acids of the fibrinogen γ chain. The α chain of fibrinogen contains an amino-terminal domain that is involved in interchain disulfide bond formation between α chains. This domain includes the amino-terminal portion of the α chain of fibrinogen from glycine, amino acid 36 to glycine, amino acid 67 of Sequence ID Number 4.

As will be evident to one skilled in the art, the hybrid proteins of the present invention may contain domains of human and other animal proteins. containing domains suitable for use in the present human and other invention from animals and the

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molecules encoding such proteins have been reported. Involucrin, loricrin, fibrinogen and fibronectin, example, have been studied in a variety of animals. DNA sequences encoding primate, canine and porcine involucrin 5 have been reported (Djian and Green, Mol. Biol. Evol. 9: 417-432, 1992; Djian and Green, Proc. Natl. Acad. Sci. USA 88: 5321-5325, 1991 and Tseng and Green, Mol. Biol. Evol. <u>7</u>: 293-302, 1990). Mehrel et al. (Cell 61: 1103-1112, have reported a DNA sequence encoding 10 loricrin. DNA sequences encoding rat and frog fibrinogen gamma chain have been reported (Haidaris and Courtney, Blood 79: 1218-1224, 1992 and Bhattacharya et al., Mol. Cell. Endocrinol. 72: 213-220, 1990; respectively). sequences encoding chicken and lamprey fibrinogen α chains 15 have been reported by Weissbach and Greininger (Proc. Natl. Acad. Sci. USA 87: 5198-5202, 1990) and Pan and Doolittle (Proc. Natl. Acad. Sci. USA 89: 2066-2070, 1992), respectively. DNA sequences encoding bovine and rat fibronectin have been reported by Petersen et 20 (Proc. Natl. Acad. Sci. USA 80: 137-141, 1983) Schwarzbauer et al., (Cell 35: 421-431, 1983). general, it is preferred to prepare proteins that contain component domains from a single species to minimize the possibility of immunogenicity. Thus, the invention provides hybrid proteins that can be used in human and veterinary medicine.

According to the present invention hybrid proteins having cross-linking and tissue activities are produced recombinantly from host transformed with a DNA construct comprising a DNA segment encoding a cross-linking domain from a first protein joined to a DNA segment encoding a tissue-binding domain from a second protein. As used within the context of the present invention, two or more DNA coding sequences are said to be joined when, as a result of in-frame fusions between the DNA coding sequences or as a result of the removal of intervening sequences by normal cellular

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processing, the DNA coding sequences can be translated into a polypeptide fusion. Unless otherwise noted, the DNA segments may be joined in any order to result in a DNA coding sequence that can be translated into a polypeptide chain. Thus, the DNA segment encoding the tissue-binding domain may be joined to the 5' or the 3' end of the DNA segment encoding the cross-linking domain. However, as will be evident to one skilled in the art, the production of hybrid proteins that additionally include domains that facilitate end-to-end cross-linking will require that the DNA segments encoding such domains be positioned at the 5' and 3' termini of the molecules.

Thus the present invention also isolated DNA molecules encoding hybrid proteins comprising a cross-linking domain from a first protein covalently linked to a tissue-binding domain from a second protein. In general, cDNA sequences are preferred for carrying out the present invention due to their lack of intervening sequences which can lead to aberrant RNA processing and reduced expression levels. DNA molecules encoding human fibronectin (Dufour et al., Exper. Cell Res. 193: 331-338, 1991) and a human fibrinogen α chain (Rixon et al., Biochemistry 22: 3250-3256, 1983) may be obtained from libraries prepared from liver cells according to standard laboratory procedures. It will be understood however, that suitable DNA sequences can also be obtained from genomic clones or can be synthesized de novo according to conventional procedures. If partial clones are obtained, it is necessary to join them in proper reading frame to produce a full length clone, using such techniques as endonuclease cleavage, ligation, and loop-out mutagenesis.

DNA sequences encoding hybrid proteins of the present invention may be prepared from cloned DNAs using conventional procedures of endonuclease cleavage, exonuclease digestion, ligation and in vitro mutagenesis. Alternatively, DNA sequences encoding the cross-linking

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and tissue-binding domains, such as those mentioned above, may be synthesized using standard laboratory techniques.

exemplary DNA molecule encoding a hybrid protein having cross-linking and tissue-binding activities may be prepared by joining a DNA segment encoding at least the cell-binding domain of fibronectin and a DNA segment encoding at least an inter-chain cross-linking domain of fibrinogen at а convenient restriction site synthetic adapters to facilitate in-frame joining of the DNA segments. Alternatively, such DNA segments encoding hybrid proteins of the present invention may be prepared by joining the two domains at a convenient restriction site followed by loop-out mutagenesis to precisely remove unnecessary sequences and directly join the DNA segment encoding the cell-binding domain of fibronectin with the DNA segment encoding the cross-linking domain of fibrinogen.

DNA segments encoding the hybrid proteins of the instant invention are inserted into DNA constructs. used within the context of the present invention, a DNA construct is understood to refer to a DNA molecule, or a clone of such a molecule, either singleor doublestranded, which has been modified through intervention to contain segments of DNA combined juxtaposed in a manner that would not otherwise exist in DNA constructs of the present invention comprise a first DNA segment encoding a hybrid protein operably to additional DNA segments required for expression of the first DNA segment. Within the context the present invention, additional DNA segments will generally include promoters and transcription terminators, and may further include enhancers and other elements.

DNA constructs may also contain DNA segments necessary to direct the secretion of a polypeptide or protein of interest. Such DNA segments may include at least one secretory signal sequence. Secretory signal sequences, also called leader sequences, prepro sequences

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and/or pre sequences, are amino acid sequences that act to direct the secretion of mature polypeptides or proteins Such sequences are characterized by a core from a cell. of hydrophobic amino acids and are typically (but not exclusively) found at the amino termini synthesized proteins. DNA segments encoding secretory signal sequences are placed in-frame and in the correct spatial relationship to the DNA segment encoding the protein of interest in order to direct the secretion of the protein. Very often the secretory peptide is cleaved from the mature protein during secretion. Such secretory peptides contain processing sites that allow cleavage of the secretory peptides from the mature proteins as they through secretory the pathway. Α preferred processing site is a dibasic cleavage site, such as that recognized by the Saccharomyces cerevisiae KEX2 gene. particularly preferred processing site is Lys-Arg processing site. Processing sites may be encoded within the secretory peptide or may be added to the peptide by, for example, in vitro mutagenesis.

Preferred secretory signals include the α factor signal sequence (pre-pro sequence: Kurjan and Herskowitz, Cell 30: 933-943, 1982; Kurjan et al., U.S. Patent No. 4,546,082; Brake, U.S. Patent No. 4,870,008), the PHO5 signal sequence (Beck et al., WO 86/00637), the BAR1 secretory signal sequence (MacKay et al., U.S. Patent No. 4,613,572; MacKay, WO 87/002670), the <u>SUC2</u> signal sequence (Carlsen et al., Molecular and Cellular Biology 3: 439-Alternately, a secretory signal sequence may 447, 1983). be synthesized according to the rules established, example, by von Heinje (European Journal of Biochemistry 133: 17-21, 1983; <u>Journal of Molecular Biology</u> 184: 99-105, 1985; Nucleic Acids Research 14: 4683-4690, 1986).

Secretory signal sequences may be used singly or may be combined. For example, a DNA segment encoding a first secretory signal sequence may be used in combination with a DNA segment encoding the third domain of barrier

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(described in U.S. Patent No. 5,037,243, which is incorporated by reference herein in its entirety). The DNA segment encoding the third domain of barrier may be positioned in proper reading frame 3' of the DNA segment of interest or 5' to the DNA segment and in proper reading frame with both the DNA segment encoding the secretory signal sequence and the DNA segment of interest.

The choice of suitable promoters, terminators and secretory signals is well within the level of ordinary skill in the art. Methods for expressing cloned genes in Saccharomyces cerevisiae are generally known in the art (see, "Gene Expression Technology," Methods in Enzymology, Vol. 185, Goeddel (ed.), Academic Press, San Diego, CA, 1990 and "Guide to Yeast Genetics and Molecular Biology," Methods in Enzymology, Guthrie and Fink (eds.), Academic Press, San Diego, CA, 1991; which are incorporated herein by reference). Transformation systems for other yeasts, including Hansenula polymorpha, Schizosaccharomyces pombe, Kluyveromyces lactis, Kluyveromyces fragilis, maydis, Pichia pastoris, Pichia guillermondil and Candida maltosa are known in the art. See, for example, Gleeson et al., J. Gen. Microbiol. 132:3459-3465, 1986 and Cregg, U.S. Patent No. 4,882,279.

Proteins of the present invention can also be 25 expressed in filamentous fungi, for example, strains of the fungi Aspergillus (McKnight et al., U.S. Patent No. 4,935,349, which is incorporated herein by reference). for transforming Acremonium chrysogenum disclosed by Sumino et al., U.S. Patent No. which is incorporated herein by reference.

Other higher eukaryotic cells may also be used as hosts, including insect cells, plant cells and avian Transformation of insect cells and production of cells. foreign proteins therein is disclosed by Guarino et al., U.S. Patent No. 5,162,222 and Bang et al., U.S. Patent No. 4,775,624, which are incorporated herein by reference. The use of Agrobacterium rhizogenes as a vector

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expressing genes in plant cells has been reviewed by Sinkar et al., <u>J. Biosci. (Bangalore)</u> 11:47-58, 1987.

Expression of cloned genes in cultured mammalian cells and in E. coli, for example, is discussed in detail in Sambrook et al. (Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor, NY, which is incorporated herein by reference). In addition E. coli. Bacillus and other genera are useful prokaryotic hosts for expressing foreign proteins. would be evident to one skilled in the art, one could express the proteins of the instant invention in other host cells such as avian, insect and plant cells using regulatory sequences, vectors and methods well established in the literature.

15 In yeast, suitable vectors for use present invention include YRp7 (Struhl et al., Proc. Natl. Acad. Sci. USA 76: 1035-1039, 1978), YEp13 (Broach et al., Gene 8: 121-133, 1979), POT vectors (Kawasaki et al, U.S. Patent No. 4,931,373, which is incorporated by reference 20 herein), pJDB249 and pJDB219 (Beggs, Nature 275:104-108, 1978) and derivatives thereof. Preferred promoters for use in yeast include promoters from yeast glycolytic genes (Hitzeman et al., <u>J. Biol. Chem. 255</u>: 12073-12080, 1980; Alber and Kawasaki, J. Mol. Appl. Genet. 1: 419-434, 1982; 25 Kawasaki, U.S. Patent No. 4,599,311) ordehydrogenase genes (Young et al., in Genetic Engineering of Microorganisms for Chemicals, Hollaender (eds.), p. 355, Plenum, New York, 1982; Ammerer, Meth. Enzymol. <u> 101</u>: 192-201, 1983). In this regard, 30 particularly preferred promoters are the TPI1 promoter (Kawasaki, U.S. Patent No. 4,599,311, 1986) and the ADH2- $\underline{4^{C}}$ promoter (Russell et al., Nature 304: 652-654, 1983; Irani and Kilgore, U.S. Patent Application Serial No. 07/631,763, CA 1,304,020 and EP 284 044, which 35 incorporated herein by reference). The expression units include a transcriptional terminator. may also Α

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preferred transcriptional terminator is the <u>TPI1</u> terminator (Alber and Kawasaki, ibid.).

Host cells containing DNA constructs of present invention are then cultured to produce the hybrid The cells are cultured according to standard methods in a culture medium containing nutrients required for growth of the particular host cells. A variety of suitable media are known in the art and generally include a carbon source, a nitrogen source, essential amino acids, vitamins, minerals and growth factors. The growth medium generally select for cells containing construct by, for example, drug selection or deficiency in essential nutrient which is complemented selectable marker on the DNA construct or co-transfected with the DNA construct.

Selection of a medium appropriate particular host cell used is within the level of ordinary skill in the Yeast cells, art. for example, preferably cultured in a chemically defined medium, comprising a non-amino acid nitrogen source, inorganic salts, vitamins and essential amino acid supplements. pH of the medium is preferably maintained at a pH greater than 2 and less than 8, preferably at pH 6.5. Methods for maintaining a stable pH include buffering and constant pH control, preferably through the addition of hydroxide or ammonium hydroxide. Preferred buffering agents include succinic acid and Bis-Tris (Sigma Chemical Co., St. Louis, MO). Yeast cells having a defect in a gene required for asparagine-linked glycosylation preferably grown in а medium containing an stabilizer. A preferred osmotic stabilizer is sorbitol supplemented into the medium at a concentration between 0.1 M and 1.5 M, preferably at 0.5 M or 1.0 M. mammalian cells are generally cultured in commercially available serum-containing or serum-free media.

The recombinant hybrid proteins expressed using the methods described herein are isolated and purified by

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conventional procedures, including separating the cells the medium centrifugation by or filtration. precipitating the proteinaceous components of the supernatant or filtrate by means of a salt, e.g. ammonium sulfate, purification by a variety of chromatographic procedures, e.g. ion exchange chromatography or affinity or chromatography, the like. Methods of protein purification are known in the art (see generally, Scopes, Protein Purification, Springer-Verlag, (1982),which is incorporated herein by reference) and may be applied to the purification of the recombinant proteins of the present invention.

The hybrid proteins of the present invention may be used as components of tissue adhesives. 15 preferred that the tissue adhesives be formulated provide a concentration of the hybrid proteins of the present invention of between about 5 mg/ml to 100 mg/ml, with concentrations in the range of 35 to 50 mg/ml being particularly preferred. As disclosed above, 20 adhesives generally contain factor XIII and thrombin. Additional components may also be included in the tissue adhesive formulations. These additional components include growth factors such as PDGF, bFGF, TGFa, or EGF and protease inhibitors, such as aprotinin, transexamic 25 acid, alpha-2 plasmin inhibitor, alpha-1-antitrypsin or the Pittsburgh mutant of alpha-1-antitrypsin (Arg-358 The tissue adhesives may alpha-1-antitrypsin). contain salts, buffering agents, reducing agents, bulking agents, and solubility enhancers. Albumin, NaCl, CaCl2, 30 citrate and phosphate buffers, for example, may included. Preferably, the tissue adhesives of the present invention are prepared as lyophilized powders, liquid concentrates of ready-to-use liquids. Lyophilized powders are preferred for ease of handling and storage.

The following examples are offered by way of illustration and not by way of limitation.

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EXAMPLES

Example 1 -Subcloning and Modification of ADH2 Promoters

An ADH2-4^C promoter was constructed as described co-pending U.S. Patent Application 07/631,763, 1,304,020 and EP 284 044, which are incorporated herein by reference. A DNA construct comprising the complete ADH2- $\underline{4^{C}}$ promoter mutagenized at the 3' end to place an Eco RI site in place of the translation start codon, designated p410-4^C (deposited with the American Type Collection (12301 Parklawn Dr., Rockville, MD 20852) under accession number 68861) was used as the source of the ADH2-4^C promoter.

A PAP-I cDNA (U.S. Patent No. 4,937,324) was 15 the ADH2-4^C promoter. joined with Plasmid comprising the 1.7 kb cDNA in pUC18, was cut with Nco I and Bam HI, and the linearized plasmid was isolated through two rounds of gel purification. The ADH2-4C promoter from p410-4° was joined to the 5' end of the PAP-20 I cDNA via an Eco RI-Nco I adapter. The 1.2 kb Bam HI-Eco RI promoter fragment from p410-4^C, Eco RI-Nco I adapter and the Nco I-Bam HI linearized pAP1.7 plasmid were The resultant plasmid was designed pPR1. ligated. presence of the correct promoter fusion was confirmed by 25 DNA sequencing.

A yeast expression vector comprising the $\underline{ADH2-4}^{\underline{C}}$ promoter, the PAP-I cDNA and the $\underline{TPI1}$ terminator was constructed. Plasmid pZUC13 (comprising the \underline{S} . cerevisiae chromosomal $\underline{LEU2}$ gene and the origin of replication from \underline{S} . cerevisiae 2 micron plasmid inserted into pUC13 and constructed in a manner analogous to pZUC12, described in published EP 195,691, using the plasmid pMT212, which is described in published EP 163 529) was cut with Bam HI. Plasmid pPR1 was digested completely digested with Bam HI and partially digested with Sac I to isolate the 2.1 kb $\underline{ADH2-4}^{\underline{C}}$ promoter-PAP-I cDNA fragment. Plasmid pTT1 (described in detail below) was digested with Sac I and

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Bam HI to isolate the 0.69 bp <u>TPI1</u> terminator fragment. The Bam HI-Sac I fragment from pPR1 and the Sac I-Bam HI fragment from pTT1 were ligated with the Bam HI-linearized pZUC13. A plasmid containing the expression unit was designated pZ3.

Example 2 - Subcloning of the TPI1 terminator

The yeast <u>TPI1</u> terminator fragment was obtained from plasmid p270 described by Murray and Kelly (U.S. Patent 4,766,073, which is incorporated by reference herein in its entirety). Plasmid p270 contains the <u>TPI1</u> terminator inserted as and Xba I-Bam HI fragment into YEp13. Alternatively, the <u>TPI1</u> terminator may be obtained from plasmid pM220 (deposited with American Type Culture Collection as an <u>E. coli</u> RR1 transformant under accession number 39853) by digesting the plasmid with Xba I, and Bam HI and purifying the <u>TPI1</u> terminator fragment (~700 bp).

The TPI1 terminator was removed from plasmid p270 as a Xba I-Bam HI fragment. This fragment was cloned into pUC19 along with another fragment containing the TPI1 promoter fused to the CAT (chloramphenicol transferase) gene to obtain a TPI1 terminator fragment with an Eco RV end. The resultant plasmid was designated The TPI1 terminator was then cut from pCAT as an Eco RV-Bam HI fragment and cloned into pIC19H (Marsh et al., Gene 32:481-486, 1984) which had been cut with the same enzymes, to obtain pTT1 (disclosed in U.S. Patent No. 4,937,324, which is incorporated herein by reference).

30 Example 3 - <u>Construction of Yeast Vectors pDPOT and pRPOT</u>

Plasmid pDPOT was derived from plasmid pCPOT (ATCC No. 39685) by replacing the 750 bp Sph I-Bam HI fragment of pCPOT containing 2 micron and pBR322 sequences with a 186 bp Sph I-Bam HI fragment derived from the pBR322 tetracycline resistance gene.

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Plasmid pRPOT was derived from plasmid pDPOT by replacing the Sph I-Bam HI fragment with a polylinker. Plasmid pDPOT was digested with Sph I and Bam HI to isolate the 10.8 kb fragment. Oligonucleotides ZC1551 and ZC1552 (Sequence ID Nos. 7 and 8) were designed to form an adapter with a Bam HI adhesive end and an Sph I adhesive end flanking Sma I, Sst I and Xho I restriction sites. Oligonucleotides ZC1551 and ZC1552 (Sequence ID Nos. 7 and 8) were kinased and annealed to form the Bam HI-Sph I adapter. The 10.8 kb pDPOT fragment was circularized by ligation with the ZC1551/ZC1552 adapter (Sequence ID Nos. 7 and 8). The resultant plasmid was termed pRPOT.

Example 4 - <u>Construction of a Fibrinogen: Fibronectin</u> Hybrid cDNA Expression Vector

A. Construction of pFN14A

Α DNA construct containing a DNA segment encoding the fibronectin cell-binding domain operably linked to the ADH2-4° promoter in plasmid pUC19 constructed. The fibronectin coding sequence was obtained ' from plasmid pFH103 (Dufour et al., Exper. Cell Res. 193: 331-338, 1991). Plasmid pFH103 was digested with Nco I and Xba to isolate the 4 kb fragment containing fibronectin coding sequence. Oligonucleotides ZC2052 and ZC2053 (Sequence ID Nos. 9 and 10) were designed to provide, upon annealing, an adapter containing a 5' Eco RI end, an internal Nco I site, a DNA segment encoding a methionine and amino acids 979 through 981 of Sequence ID Number 2 and a 3' Nco I adhesive end that destroys the Nco I site. Oligonucleotides ZC2052 ZC2053 (Sequence ID Nos. 9 and 10) were annealed ligated with the 4 kb Nco I-Xba I fibronectin fragment into Eco RI-Xba I linearized pUC19. The resultant plasmid was designated pFN4.

Plasmid pFN4 was digested with Hind III and Apa I to isolate the 3.3 kb fibronectin fragment.
Oligonucleotides ZC2493 and ZC2491 (Sequence ID Nos. 12

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and 11) were designed to provide, when annealed, an Apa I-Xba I adapter encoding the amino acids Pro and Phe followed by a stop codon. The oligonucleotides were annealed and combined with the 3.3 kb Hind III-Apa I fragment and Hind III-Xba I linearized pUC19 to form plasmid pFN7. Plasmid pFN7 comprises a DNA segment encoding amino acids 1273-2186 of Sequence ID Number 2 followed by an in-frame stop codon.

The <u>ADH2-4^C</u> promoter was joined to the 5' end of the fibronectin cDNA in plasmid pFN5. Plasmid pFN4 was digested with Nco I and Hind III to isolate the 0.89 kb fibronectin coding sequence. Plasmid pZ3 (described in detail above) was digested with Bam HI and Nco I to isolate the 1.25 kb <u>ADH2-4^C</u> promoter fragment. The 1.25 kb Bam HI-Nco I promoter fragment and the Nco I-Hind III fibronectin coding sequence fragment were ligated to Bam HI-Hind III linearized pUC19 to form plasmid pFN5.

Plasmid pFN5 was digested with Bam HI and Hind III to isolate the 2.1 kb promoter-fibronectin fragment. Plasmid pFN7 was digested with Hind III and Xba I to isolate the 2.8 kb fibronectin fragment that was modified to encode a stop codon following the Pro-Phe sequence. The TPI1 terminator sequence was obtained from pTT1 as a 0.7 kb Xba I-Sal I fragment. The 2.1 kb Bam HI-Hind III promoter-fibronectin fragment, the 2.8 kb Hind III-Xba I fibronectin fragment and the 0.7 kb TPI1 terminator fragment were joined in a four-part ligation with Bam HIlinearized pRPOT. A plasmid containing fibronectin expression unit in the pRPOT was designated pR1.

The original clone pFH103 contained a frameshift mutation in the EIIIB region of the fibronectin cDNA. The mutation was corrected by the replacement of the region with an analogous region from the plasmid pFHA3 (obtained from Jean Paul Thiery, Laboratoire de Physiopathologie du Developpement, CNRS URA 1337, Ecole Normale Superiure, 46 rue d'Ulm, 75230 Paris Cedex 05,

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France). Plasmid pFH∆3 was derived from pFH103 excising the 3211 qd Xba I-Asp 718I fragment of fibronectin, blunting of the resultant adhesive ends and Plasmid pFHA3 contains a DNA segment encoding religating. the signal and propeptides, the first three and one half type I repeats, and the carboxy-terminal half of human fibronectin from the middle of the EIIIB segment.

Plasmid pR1 was digested with Bam HI and Kpn I to 2.2 kb promoter-fibronectin the Plasmid pFHA3 was digested with Kpn I and Apa I to isolate the internal fibronectin fragment that corrects the frameshift mutation present in the parent cDNA from pFH103. Plasmid pR1 was digested with Apa I and Bam HI to isolate the TPI1 terminator fragment. The 2.2 kb Bam HI-Kpn I promoter-fibronectin fragment, the 2.75 kb Kpn I-Apa I internal fibronectin fragment and the 0.69 kb Apa I-Bam HI terminator fragment were in joined а four-part ligation with Bam HI-linearized pDPOT. The resulting construction was designated pD32.

A DNA segment encoding the <u>ADH2-4</u>^C promoter and initiation methionine from plasmid pD32 was subcloned into pIC19H (Marsh et al., <u>Gene 32</u>:481-486, 1984) as a 1.25 kb Bam HI-Nco I fragment. Plasmid pD32 was also digested with Nco I and Bgl II to isolate the 3 kb fibronectin cDNA fragment encoding amino acids 979-1972 of Sequence ID Number 2. The 1.25 kb Bam HI-Nco I fragment and the Nco I-Bgl II fragment were ligated with Bam HI-linearized pIC19H. A plasmid containing a Bam HI site proximal to the <u>ADH2-4</u>^C promoter was designated pFN14A.

B. - Construction of Plasmid pD38

An expression vector comprising a DNA segment encoding a fibronectin-fibrinogen hybrid protein operably linked to the $\underline{ADH2-4}^{C}$ promoter and the TPI1 terminator was constructed. To assemble the DNA sequence encoding the hybrid protein, a DNA segment encoding approximately the

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carboxy-terminal 409 amino acids of the α chain of fibrinogen was first subcloned.

A fibrinogen α chain cDNA was obtained from Dominic W. Chung (Department of Biochemistry, University of Washington, Seattle, WA) in plasmid pHI α 3 (Rixon et al., <u>Biochemistry 22</u>: 3250-3256, 1983). Sequence analysis of the cDNA insert in plasmid pHI α -3 revealed a deletion of codons 1348-1350 of the published sequence resulting in the deletion of Serine, amino acid 417.

The DNA segment encoding the carboxy-terminus of the fibrinogen α chain was subcloned into plasmid pUC19. Plasmid pHI α -3 was digested with Asp 718 and Ssp I to isolate the approximately 2 kb fragment encoding the carboxy-terminus of the fibrinogen a chain from amino acid 244 to amino acid 643 and some 3' untranslated sequence of Sequence ID Number 4. Plasmid pTT1 was digested with Eco RV and Sal I to isolate the approximately 700 bp TPI1 The 2 kb fibrinogen α chain sequence terminator fragment. and the TPI1 terminator sequence were ligated with pUC19 that had been linearized with Asp 718 and Sal I. ligation mixture was transformed into E. coli, and plasmid DNA was prepared and analyzed by restriction endonuclease and DNA sequence analysis. DNA sequence analysis of a candidate clone revealed that the Sal I site joining the TPI1 terminator sequence and the pUC19 polylinker site was not present. Plasmid DNA from the candidate clone was digested with Asp 718 and Bam HI to liberate approximately 1.9 kb fibrinogen-TPI1 terminator fragment.

To join the fibronectin coding sequence with the fibrinogen α chain sequence, synthetic oligonucleotides were synthesized to provide, when annealed, a Sal I-Asp 718 adapter encoding an internal Afl II restriction site, and a sequence encoding amino acids 1886 through 1903 of fibronectin (Sequence ID Number 2), a glycine residue and amino acids 235 through 243 of the fibrinogen α chain (Sequence ID Number 4). Oligonucleotides ZC3521 and ZC3522 (Sequence ID Nos. 13 and 14) were annealed. The

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1.9 kb Asp 718-Bam HI fibrinogen-<u>TPI1</u> terminator fragment and the Sal I-Asp 718 ZC3521/ZC3522 adapter (Sequence ID Nos. 13 and 14) were ligated with pUC19 that had been linearized with Sal I and Bam HI. The resultant plasmid was designated pFG4.

The DNA segment encoding the fibronectinfibrinogen α chain sequence in plasmid pFG4 was joined the DNA segment encoding the amino-terminal fibronectin sequence (from amino acid 989 to amino acid Sequence ID Number 2) in plasmid construct plasmid pD37. Plasmid pFN14A was digested with Bgl II and Afl II to isolate the approximately 3.9 kb ADH2-4^C promoter-fibronectin fragment. Plasmid pFG4 was digested with Afl ΙI and Bam HI to isolate approximately 2 kb fibronectin-fibrinogen-TPI1 terminator The 3.9 kb Bgl II-Afl II fragment and the 2 kb Afl II-Bam HI fragment were ligated with Bam HI-linearized A plasmid with the expression unit inserted with the direction of transcription in the same direction as the POT1 gene in the pDPOT vector was designated pD37.

To place the expression unit present in pD37 in opposite orientation, such that the direction transcription of the expression unit was in the opposite direction to that of the POT1 gene, plasmid pD37 was digested with Nco I and Xba I to isolate the approximately 4 kb fibronectin-fibrinogen α chain fragment. pFN14A was digested with Bam HI and Nco I to isolate the approximately 1.3 kb $\underline{ADH2-4}^{\underline{C}}$ promoter fragment. pTT1 was digested with Bam HI and Xba I to isolate the approximately 700 bp TPI1 terminator fragment. HI-Nco I ADH2-4^C promoter fragment, the Nco I-Xba I fibronectin-fibrinogen α chain fragment and the Xba I-Bam TPI1 terminator fragment were ligated with Bam HIlinearized pDPOT that had been treated with calf alkaline phosphatase to prevent recircularization. containing the expression unit in the opposite orientation relative to the POT1 gene was designated pD38.

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nucleotide sequence and deduced amino acid sequence of the DNA segment encoding the fibronectin-fibrinogen hybrid of plasmid pD38 is shown in Sequence ID Number 5. Plasmid pD38 was deposited on December 15, 1992 with the American Type Culture Collection (12301 Parklawn Drive, Rockville, MD) as an E. coli transformant.

Example 5 - Expression of a Fibronectin-Fibrinogen Hybrid Protein in Yeast

10 Plasmid pD38 was transformed into Saccharomyces cerevisiae host strain ZM118 $(MATa/MAT\alpha)$ ura3/ura3 ∆tpi1::URA3/∆tpi1::URA3 <u>leu2-3,112/leu2-3,112</u> bar1/bar1 pep4::URA3/pep4::URA3 [cir^O]) using essentially the method described by Hinnen et al. (Proc. Natl. Acad. 15 Sci. USA 75: 1929-1933, 1978). Transformants selected for their ability to grow on medium containing glucose as the sole carbon source.

The ZM118[pD38] transformant was scaled up in a fermenter to facilitate purification of the hybrid protein. A single ZM118[pD38] colony was selected from a YEPD + Ade + Leu plate (Table 1) and inoculated into -LeuTrpThrD medium (Table 1). The culture was incubated for approximately 52 hours after which the cells were harvested. The cells were washed in T.E. buffer (Sambrook et al., ibid.), resuspended in T.E. buffer + 30% glycerol, and aliquotted into 1 ml seed vials. seed vials were stored at -80°C. One seed vial was used to inoculate 100 ml of YEPD + Ade + Leu (Table 1). culture was grown for approximately 28 hours to a final The 100 ml culture of ZM118[pD38] was A₆₆₀ of 7.7. inoculated into a 10 liter fermenter with a final volume of 6.0 liters of medium containing 10 g/L $(NH_4)_2SO_4$, 5 g/L KH₂PO₄, 5 g/L MgSO₄ 7H₂O, 1 g/L NaCl, 0.5 g/L CaCl₂ 2H₂O, 3.68 g/L A.A.I. (Table 1), 4.2 g/L citric acid, 60 g/L glucose, 10 ml/L Trace Metal Solution (Table 1), 0.4 ml/L (Polypropylene glycol, MW 2025, Union Carbide Corp, Danbury, CT) that had been pH adjusted to pH 5.0

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with NaOH. In addition to the inoculation culture, 30 ml of Vitamin solution was added (Table 1). The culture was grown for 23 hours at 30° C with the addition of 2 M NaOH to maintain pH of approximately 5.

<u>Table</u> 1 <u>Media Recipes</u>

5	-LeuThrTrp Amino Acid Mixture	
		adenine
	3 g	L-arginine
	_	L-aspartic acid
	-	L-histidine free base
10	6 g	L-isoleucine
	4 g	L-lysine-mono hydrochloride
		L-methionine
	6 g	L-phenylalanine
	5 g	L-serine
15	5 g	L-tyrosine
	4 g	uracil
	6 g	L-valine
		Mix all the ingredients and grind with
20	a mo	rtar and pestle until the mixture is finely
	grou	nd.
		<u>TrpThrD</u>
	20 g	-
25	6.7	•
		acids (DIFCO Laboratories, Detroit,
		MI)
	0.6	-
	18 g	Agar
30		
		Mix all the ingredients in distilled
	water	The desired was a second of the second of th
		ter. Autoclave 15 minutes. Pour plates and
	allo	v to solidify.

Table 1 continued

	YEPD + Ad	le + Leu Plates
	20 g	glucose
	20 g	Bacto Peptone (DIFCO Laboratories)
5	10 g	Bacto Yeast Extract (DIFCO
		Laboratories)
	18 g	agar
	4 ml	1% adenine
	8 ml	1% L-leucine
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		Mix all ingredients in distilled
	water, ar	nd bring to a final volume of 1 liter.
	Autoclave	25 minutes and pour plates.
15	YEPD + Ad	e + Leu Medium
	20 g	glucose
	20 g	Bacto Peptone (DIFCO Laboratories)
	10 g	Bacto Yeast Extract (DIFCO
		Laboratories)
20	4 ml	1% adenine
	8 ml	1% L-leucine
		Mix all ingredients in distilled
25		d bring to a final volume of 1 liter.
25	Autoclave	25 minutes.

Table 1 continued

	A.A.I.	
	4.0 g	adenine
	5.0 g	L-alanine
5	2.0 g	L-arginine
	5.0 g	L-asparagine
	5.0 g	L-aspartic acid
	5.0 g	L-cysteine
	5.0 g	L-glutamine
10	5.0 g	L-glutamic acid
	5.0 g	L-glycine
	8.0 g	L-histidine
	5.0 g	L-isoleucine
	3.0 g	L-lysine-mono hydrochloride
15	2.0 g	L-methionine
	5.0 g	L-phenylalanine
	5.0 g	L-proline
	5.0 g	L-serine
	5.0 g	L-threonine
20	2.0 g	L-tryptophan
	3.0 g	L-tyrosine
	3.0 g	uracil
	5.0 g	L-valine
25		Mix all the ingredients and gri

Mix all the ingredients and grind with a mortar and pestle until the mixture is finely ground. Store at room temperature.

to

Table 1 continued

	Trace Metal Solution		
	0.68 g	ZnCl ₂	
	5.4 g	FeCl ₃ ·6H ₂ O	
5	1.91 g	MnCl ₂ ·4H ₂ O	
	0.22 g	CuSO ₄ · 5H ₂ O	
	0.258 g	CoCl ₂	
	0.062 g	н ₃ во ₃	
	0.002 g	$(NH_4)_6Mo_2O_2$	
10	0.002 g	KI	
	10.0 ml 3	7% HCl	
		Dissolve solids in water and bring	
	a final vo	olume of 1 liter.	
15			
	<u>Vitamin So</u>	<u>olution</u>	
	25 mg	d-biotin	
	400 mg	thiamine	
	400 mg	pyridoxine	
20	7.5 a	meso-inositol	

400 mg thiamine
400 mg pyridoxine
20 7.5 g meso-inositol
7.5 g Ca pantothenate
300 mg niacinamide
50 mg folic acid
100 mg riboflavin
25 500 mg choline

Dissolve solids in water and bring to a final volume of 1 liter.

A 60 liter fermenter with a final volume of 50 liters of medium containing 60 g/L yeast extract (Universal Foods, Milwaukee, WI), 2.5 g/L MgSO₄·7H₂O (Mallinkrodt Inc., St. Louis, MO), 1 g/L CaCl₂·2H₂O (Mallinkrodt, Inc.), 1 g/L KCl (Mallinkrodt, Inc.), 10 ml/L of Trace Metal Solution (Table 1), 0.5 ml/L PPG-2025 (Union Carbide) that had been adjusted to a pH of 5.0 with

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H₃PO₄ was prepared, and the medium was sterilized. sterilization, 5.0 liters of the 23 hour fermentation culture and 500 ml of Vitamin Solution (Table 1) were inoculated into the medium. During the fermentation, a solution of 50% glucose, 5% $(NH_4)_2SO_4$, 0.05% citric acid was fed into the fermenter at a rate of 150 ml/hour, and pH was maintained at approximately pH 5 addition of 2 M NH4OH. PPG-2025 was added as needed to foaming. Αt approximately 49 hours inoculation, an ethanol feed was begun by the addition of ethanol to the fermenter at a rate of 150 ml/min. culture was grown for a total of 67.25 hours at 30°C.

At the end of the fermentation, 50 liters of the culture was diluted to 100 liters with water. were removed from the spent medium by centrifuging 50 liters at a time through a Westfalia CSA 19 centrifuge (Westfalia, Oelde, Germany) at a flow liters/min. The cells were rinsed with water. centrifugation, approximately 20 liters of cell containing approximately 35% cells was obtained. were added to the slurry to achieve a final concentration of the following salts: 50 mM NaCl, 10 mM Na₂HPO₄, 5 mM The cell slurry was passed through a Dynomill bead mill using 0.5 mm lead-free glass beads (Willy A Bachofen AG MashinenFabrik, Basle, Switzerland) at a rate of 4 liters per minute. The Dynomill was rinsed with Lysis buffer (50 mM NaCl, 10 mM Na₂HPO₄, 5 mM EDTA, pH 7.2) to a final volume of 80 liters. The final slurry had a pH of 6.8, a temperature of approximately 10°C and conductivity of 5 ms/cm.

The cell slurry was subjected to centrifugation as described above, and the cell pellet was rinsed with lysis buffer. After centrifugation approximately 20 liters of cell slurry was obtained. The cell slurry was extracted by first adjusting the concentration of the cell debris to approximately 40-50% with lysis buffer. Solid urea, NaCl and EDTA were added to the cell slurry to

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achieve a final concentration of approximately 8 M urea, NaCl EDTA. and 10 mM The approximate concentrations were obtained by the addition of 450 g/L of urea, 18 g/L of NaCl and 4.2 g/L of EDTA. The cell slurry was adjusted to pH 7.8 with 0.5 M NaOH. The solids were dissolved into the slurry and the pellets were extracted for a total of 50 minutes. Following extraction, the mixture was diluted 1 to 4 with water, adjusted to a conductivity of 12.5 ms/cm with NaCl and adjusted to a pH of 9.5 with 0.5 M NaOH.

The extracted slurry was centrifuged as described above with the lysis buffer rinse. The the supernatant was adjusted to pH 9.5 with 0.5 M NaOH. The supernatant was analyzed by SDS polyacrylamide gel electrophoresis (SDS-PAGE) using the PHAST Separation and Control Unit (Pharmacia LKB Biotechnology Piscataway, NJ), and the protein was visualized using Coomassie Blue staining. A 2 liter Q-sepharose column (Pharmacia) was equilibrated at 5 liters/hour with successive washes of the following solutions: 8 liters of 3 M urea, 1 M NaCl, 50 mM glycine, pH 11.5; 5 liters of 0.5 M NaOH; 1.5 liters of water; 5 liters of 0.1 M HCl; and 6.0 liters of Wash buffer (50 mM glycine, 90 mM NaCl, Нq with a conductivity of 12.5 ms/cm). supernatant (110 liters) was then applied to the column at 5 liters per hour.

The column ran dry after loading The gel was resuspended in Wash buffer and supernatant. repacked. The repacked column was washed with 4 liters of 50 mM glycine, 90 mM NaCl, 5 mM EDTA, pH 10.0. material was eluted with elution buffer (50 mM glycine, 5 mM EDTA (pH 9.9) with a final concentration of NaCl giving a conductivity of 30.2 cm/ms (approximately 270 mM NaCl)) at 100 ml per minute. The approximately 600 ml fractions were collected after the conductivity of the reached the conductivity of the elution buffer.

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were analyzed by SDS-PAGE analysis as described above and fractions 1 through 10 were pooled.

The pooled fractions were then applied to a 2 liter phenyl Sepharose column (Pharmacia) that had been equilibrated by successive washes at 5 liters per hour with the following solutions: 3 liters of 0.5 M NaOH; 3 liters of water; 3 liters of 2 M urea, 50 mM glycine, pH 10.5; 1.5 liters of water; 3 liters of 0.1 M HCl; and 3 liters of Equilibration buffer (50 mM glycine, 2.5 M NaCl, 2 mM EDTA (pH 10.0) with a conductivity of 180 ms/cm). The pooled peak fractions, which had been adjusted to a conductivity of 180 ms/cm with NaCl and a pH of 10.0 with 0.5 M NaOH, were loaded onto the phenyl sepharose column. Following the loading of the peak fractions, the column was washed with Equilibration buffer. The column was eluted with 6 liters of 50 mM glycine, 2 mM EDTA (pH 10.25) with a NaCl concentration giving the solution a The conductivity of the eluant conductivity of 96 ms/cm. was measured throughout the elution. The conductivity of the eluant upon starting the elution was 180 ms/cm. the third fraction, the conductivity of the eluant dropped to 96 ms/cm. At this point, the elution buffer was changed to a buffer having the conductivity of 42 ms/cm. The eluant was collected through fraction number 8.

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Example 6 -Cross-Linking Assay Using the Fibrinogen-Fibronectin Protein

The ability of the purified fibrinogenfibronectin hybrid protein to form transglutminasecatalyzed interchain cross links was assessed. transglutaminase activity was provided by the addition of recombinant factor XIII and thrombin or by the addition of recombinant factor XIIIa.

10 Α. Preparation of Factor XIII

Recombinant factor XIII was prepared essentially as described in co-pending U.S. Patent Application No. 07/927,196, which is incorporated by reference herein in its entirety. Briefly, factor XIII was isolated from a 15 strain of the yeast Saccharomyces cerevisiae that had been transformed with an expression vector capable of directing the expression of factor XIII. The factor XIII-producing cells were harvested and lysed, and a cleared lysate was prepared. The lysate was fractionated by anion exchange chromatography at neutral to slightly alkaline pH using a column of derivatized agarose, such as DEAE FAST-FLOW SEPHAROSE (Pharmacia LKB Biotechnology, Piscataway, NJ) or Factor XIII was then precipitated from the the like. column eluate by concentrating the eluate and adjusting the pH to between 5.2 and 5.5, such as by diafiltration against ammonium succinate buffer. The precipitate was then dissolved and further purified using conventional chromatographic techniques, such as gel filtration hydrophobic interaction chromatography. The purified factor XIII was dialyzed, filtered, aliquotted lyophilized. The factor XIIIa content was determined (Bishop et al., Biochemistry 29: 1861-1869, 1990, which is incorporated by reference herein in its entirety) fluorometric assay of the dissolved, thrombin-activated material.

Factor XIII was activated to factor XIIIa by adding 2 U of thrombin per 100 mg of factor XIII.

factor XIII was dissolved in buffer (20 mM sodium borate (pH 8.3), 1 mM $CaCl_2$). The thrombin was added, and the reaction was incubated at room temperature for twenty minutes.

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B. Cross-Linking Assays

The level of cross-linking between the hybrid proteins was measured as a rise in the absorbance at 350 nm over time in reaction mixtures containing the hybrid protein, factor XIII and thrombin or the hybrid protein factor XIIIa. Control reactions were containing factor XIII and thrombin or factor XIIIa alone. Cross-linking reactions were carried out in 1 ml cuvettes. For cross-linking reactions containing factor XIII thrombin, each reaction mixture was set up by placing 110 μl containing 40 Units of factor XIII, 36.7 μl containing 13 Units of factor XIII or 12.2 μ l containing 4 Units of factor XIII (described above) in one corner of the cuvette and 20 µl containing 4 Units of thrombin (Sigma) in the opposite corner such that the solutions were not mixed. The reaction was initiated by the addition of 1 ml of 2 mg/ml hybrid protein in buffer (10 mM Tris (pH 7.6), 20 mM sodium borate, 140 mM NaCl, 10 mM CaCl₂). The absorbance of each reaction was read at 350 nm with the addition of protein being the first absorbance point. For crosslinking reactions containing factor XIIIa, each reaction was set up by placing 110 μ l containing 40 Units of factor XIIIa, 36.7 µl containing 13 Units of factor XIIIa or 12.2 μ l containing 4 Units of factor XIIIa in the cuvette and adding 1 ml of 2 mg/ml hybrid in buffer (10 mM Tris (pH 7.6), 140 mM NaCl, 10 mM CaCl₂). The absorbance of the solution was read at 350 nm as described above. of the data generated from the absorbance time courses showed a sharp increase in absorbance in the presence of hybrid protein and the active transglutaminase relative to the rise in absorbance in the absence of hybrid protein

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(Figures 2-5). The results indicated that the hybrid protein is capable of transglutaminase-induced crosslinking.

From the foregoing it will be appreciated that, although specific embodiments of the invention have been described herein for the purpose of illustration, various modifications may be made without deviation from the spirit and scope of the invention. Accordingly, the invention is not to be limited except as by the following claims.

SEQUENCE LISTING

- (1) GENERAL INFORMATION:
 - (i) APPLICANT: Irani, Meher H.
 - (ii) TITLE OF INVENTION: HYBRID CROSS-LINKING PROTEINS
 - (iii) NUMBER OF SEQUENCES: 14
 - (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: ZymoGenetics, Inc.
 - (B) STREET: 4225 Roosevelt Way, N.E.
 - (C) CITY: Seattle
 - (D) STATE: WA
 - (E) COUNTRY: USA
 - (F) ZIP: 98105
 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk

 - (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
 - (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: WO
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
 - (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 07/998,271
 - (B) FILING DATE: 31-DEC-1992
 - (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Parker, Gary E
 - (B) REGISTRATION NUMBER: 31-648
 - (C) REFERENCE/DOCKET NUMBER: 92-26PC
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: 206-547-8080 ext 322
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- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 7803 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS

(B) LOCATION: 6..7346

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

TCA	AC A'	TG C et Lo 1	TT A	GG G(rg G	GT CO	CG G(ro G	GG CO	CC G(ro G	GG C'	eu L	TG C' eu Lo 10	TG C' eu L	TG G eu A	CC G la V	TC al	47
CTG Leu 15	TGC Cys	CTG Leu	GGG Gly	ACA Thr	GCG Ala 20	GTG Val	CCC Pro	TCC Ser	ACG Thr	GGA Gly 25	GCC Ala	TCG Ser	AAG Lys	AGC Ser	AAG Lys 30	95
AGG Arg	CAG Gln	GCT Ala	CAG Gln	CAA Gln 35	ATG Met	GTT Val	CAG Gln	CCC Pro	CAG Gln 40	TCC Ser	CCG Pro	GTG Val	GCT Ala	GTC Val 45	AGT Ser	143
		AAG Lys														191
		TGG Trp 65														239
Tyr	GGA Gly 80	GGA Gly	AGC Ser	CGA Arg	GGT Gly	TTT Phe 85	AAC Asn	TGC Cys	GAA Glu	AGT Ser	AAA Lys 90	CCT Pro	GAA Glu	GCT Ala	GAA Glu	287
GAG Glu 95	ACT Thr	TGC Cys	TTT Phe	GAC Asp	AAG Lys 100	TAC Tyr	ACT Thr	GGG Gly	AAC Asn	ACT Thr 105	TAC Tyr	CGA Arg	GTG Val	GGT Gly	GAC Asp 110	335
ACT Thr	TAT Tyr	GAG Glu	CGT Arg	CCT Pro 115	AAA Lys	GAC Asp	TCC Ser	ATG Met	ATC Ile 120	TGG Trp	GAC Asp	TGT Cys	ACC Thr	TGC Cys 125	ATC Ile	383
GGG Gly	GCT Ala	GGG Gly	CGA Arg 130	GGG Gly	AGA Arg	ATA Ile	AGC Ser	TGT Cys 135	ACC Thr	ATC Ile	GCA Ala	AAC Asn	CGC Arg 140	TGC Cys	CAT His	431
GAA Glu	GGG Gly	GGT Gly 145	CAG Gln	TCC Ser	TAC Tyr	AAG Lys	ATT Ile 150	GGT Gly	GAC Asp	ACC Thr	TGG Trp	AGG Arg 155	AGA Arg	CCA Pro	CAT His	479
GAG Glu	ACT Thr 160	GGT Gly	GGT Gly	TAC Tyr	ATG Met	TTA Leu 165	GAG Glu	TGT Cys	GTG Val	TGT Cys	CTT Leu 170	GGT Gly	AAT Asn	GGA Gly	AAA Lys	527
GGA Gly 175	GAA Glu	TGG Trp	ACC Thr	TGC Cys	AAG Lys 180	CCC Pro	ATA Ile	GCT Ala	GAG Glu	AAG Lys 185	TGT Cys	TTT Phe	GAT Asp	CAT His	GCT Ala 190	5 75
		ACT Thr														623

GGC Gly	TGG Trp	ATG Met	ATG Met 210	GTA Val	GAT Asp	TGT Cys	ACT Thr	TGC Cys 215	CTG Leu	GGA Gly	GAA Glu	GGC Gly	AGC Ser 220	GGA Gly	CGC Arg	671
			ACT Thr												ACA Thr	719
TCC Ser	TAT Tyr 240	AGA Arg	ATT Ile	GGA Gly	GAC Asp	ACC Thr 245	TGG Trp	AGC Ser	AAG Lys	AAG Lys	GAT Asp 250	AAT Asn	CGA Arg	GGA Gly	AAC Asn	767
CTG Leu 255	CTC Leu	CAG Gln	TGC Cys	ATC Ile	TGC Cys 260	ACA Thr	GGC Gly	AAC Asn	GGC Gly	CGA Arg 265	GGA Gly	GAG Glu	TGG Trp	AAG Lys	TGT Cys 270	815
GAG Glu	AGG Arg	CAC His	ACC Thr	TCT Ser 275	GTG Val	CAG Gln	ACC Thr	ACA Thr	TCG Ser 280	AGC Ser	GGA Gly	TCT Ser	GGC Gly	CCC Pro 285	TTC Phe	863
ACC Thr	GAT Asp	GTT Val	CGT Arg 290	GCA Ala	GCT Ala	GTT Val	TAC Tyr	CAA Gln 295	CCG Pro	CAG Gln	CCT Pro	CAC His	CCC Pro 300	CAG Gln	CCT Pro	911
CCT Pro	CCC Pro	TAT Tyr 305	GGC Gly	CAC His	TGT Cys	GTC Val	ACA Thr 310	GAC Asp	AGT Ser	GGT Gly	GTG Val	GTC Val 315	TAC Tyr	TCT Ser	GTG Val	959
GGG Gly	ATG Met 320	CAG Gln	TGG Trp	TTG Leu	AAG Lys	ACA Thr 325	CAA Gln	GGA Gly	AAT Asn	AAG Lys	CAA Gln 330	ATG Met	CTT Leu	TGC Cys	ACG Thr	1007
TGC Cys 335	CTG Leu	GGC Gly	AAC Asn	GGA Gly	GTC Val 340	AGC Ser	TGC Cys	CAA Gln	GAG Glu	ACA Thr 345	GCT Ala	GTA Val	ACC Thr	CAG Gln	ACT Thr 350	1055
TAC Tyr	GGT Gly	GGC Gly	AAC Asn	TTA Leu 355	AAT Asn	GGA Gly	GAG Glu	CCA Pro	TGT Cys 360	GTC Val	TTA Leu	CCA Pro	TTC Phe	ACC Thr 365	TAC Tyr	1103
AAT Asn	GGC Gly	AGG Arg	ACG Thr 370	TTC Phe	TAC Tyr	TCC Ser	TGC Cys	ACC Thr 375	ACG Thr	GAA Glu	GGG Gly	CGA Arg	CAG Gln 380	GAC Asp	GGA Gly	1151
CAT His	CTT Leu	TGG Trp 385	TGC Cys	AGC Ser	ACA Thr	ACT Thr	TCG Ser 390	AAT Asn	TAT Tyr	GAG Glu	CAG Gln	GAC Asp 395	CAG Gln	AAA Lys	TAC Tyr	1199
TCT Ser	TTC. Phe 400	TGC Cys	ACA Thr	GAC Asp	CAC His	ACT Thr 405	GTT Val	TTG Leu	GTT Val	CAG Gln	ACT Thr 410	CAA G1n	GGA Gly	GGA Gly	AAT Asn	1247
TCC Ser 415	AAT Asn	GGT Gly	GCC Ala	TTG Leu	TGC Cys 420	CAC His	TTC Phe	CCC Pro	TTC Phe	CTA Leu 425	TAC Tyr	AAC Asn	AAC Asn	CAC His	AAT Asn 430	1295

			GAG Glu						1343
			GAT Asp						1391
			ATC Ile						1439
			GAT Asp 485						1487
			AAT Asn						1535
			CAG Gln						1583
			AAG Lys						1631
			GGT Gly						1679
			GAG Glu 565						1727
			CAT His						1775
			TGG Trp						1823
			GAA Glu						1871
			CAG Gln						1919

TCC Ser	AAG Lys 640	Tyr	ATT 11e	CTC Leu	AGG Arg	TGG Trp 645	Arg	CCT Pro	AAA Lys	AAT Asn	TCT Ser 650	· Val	GGC Gly	CG7	TGG Trp	19	967
AAG Lys 655	Glu	GCT Ala	ACC Thr	: ATA	CCA Pro 660	Gly	CAC His	TTA Leu	AAC Asn	TCC Ser 665	Tyr	ACC Thr	ATC Ile	: AAA : Lys	GGC Gly 670	20	015
CTG Leu	AAG Lys	CCT Pro	GGT Gly	GTG Val 675	Val	TAC Tyr	GAG Glu	GGC Gly	CAG Gln 680	Leu	ATC Ile	AGC Ser	ATC	CAG Gln 685	CAG Gln	20	063
TAC Tyr	GGC Gly	CAC His	CAA Gln 690	Glu	GTG Val	ACT Thr	CGC Arg	TTT Phe 695	GAC Asp	TTC Phe	ACC Thr	ACC Thr	ACC Thr 700	Ser	ACC Thr	21	111
AGC Ser	ACA Thr	CCT Pro 705	Val	ACC Thr	AGC Ser	AAC Asn	ACC Thr 710	GTG Val	ACA Thr	GGA Gly	GAG Glu	ACG Thr 715	ACT Thr	CCC Pro	TTT Phe	21	159
TCT Ser	CCT Pro 720	Leu	GTG Val	GCC Ala	ACT Thr	TCT Ser 725	GAA Glu	TCT Ser	GTG Val	ACC Thr	GAA G1u 730	Ile	ACA Thr	GCC Ala	AGT Ser	22	207
AGC Ser 735	TTT Phe	GTG Val	GTC Val	TCC Ser	TGG Trp 740	GTC Val	TCA Ser	GCT Ala	TCC Ser	GAC Asp 745	ACC Thr	GTG Val	TCG Ser	GGA Gly	TTC Phe 750	22	255
CGG Arg	GTG Val	GAA Glu	TAT Tyr	GAG Glu 755	CTG Leu	AGT Ser	GAG Glu	GAG Glu	GGA Gly 760	GAT Asp	GAG Glu	CCA Pro	CAG Gln	TAC Tyr 765	CTG Leu	23	103
GAT Asp	CTT Leu	CCA Pro	AGC Ser 770	ACA Thr	GCC Ala	ACT Thr	TCT Ser	GTG Val 775	AAC Asn	ATC Ile	CCT Pro	GAC Asp	CTG Leu 780	CTT Leu	CCT Pro	23	51
GGC Gly	CGA Arg	AAA Lys 785	TAC Tyr	ATT Ile	GTA Val	AAT Asn	GTC Val 790	TAT Tyr	CAG Gln	ATA Ile	TCT Ser	GAG G1u 7 9 5	GAT Asp	GGG Gly	GAG Glu	23	99
CAG Gln	AGT Ser 800	TTG Leu	ATC Ile	CTG Leu	TCT Ser	ACT Thr 805	TCA Ser	CAA Gln	ACA Thr	ACA Thr	GCG Ala 810	CCT Pro	GAT Asp	GCC Ala	CCT Pro	24	47
CCT Pro 815	GAC Asp	CCG Pro	ACT Thr	GTG Val	GAC Asp 820	CAA Gln	GTT Val	GAT Asp	GAC Asp	ACC Thr 825	TCA Ser	ATT Ile	GTT Val	GTT Val	CGC Arg 830	24	95
TGG Trp	AGC Ser	AGA Arg	CCC Pro	CAG Gln 835	GCT Ala	CCC Pro	ATC Ile	Thr	GGG Gly 840	TAC Tyr	AGA Arg	ATA Ile	GTC Val	TAT Tyr 845	TCG Ser	25	43
CCA Pro	TCA Ser	GTA Val	GAA Glu 850	GGT Gly	AGC Ser	AGC Ser	ACA Thr	GAA Glu 855	CTC Leu	AAC Asn	CTT Leu	CCT Pro	GAA Glu 860	ACT Thr	GCA Ala	259	91

AAC Asn	TCC Ser	GTC Val 865	ACC Thr	CTC Leu	AGT Ser	GAC Asp	TTG Leu 870	CAA Gln	CCT Pro	GGT Gly	GTT Val	CAG Gln 875	TAT Tyr	AAC Asn	ATC Ile	2639
ACT Thr	ATC Ile 880	TAT Tyr	GCT Ala	GTG Val	GAA Glu	GAA Glu 885	AAT Asn	CAA Gln	GAA Glu	AGT Ser	ACA Thr 890	CCT Pro	GTT Val	GTC Val	ATT Ile	2 <u>687</u>
CAA G1n 895	CAA Gln	GAA Glu	ACC Thr	ACT Thr	GGC Gly 900	ACC Thr	CCA Pro	CGC Arg	TCA Ser	GAT Asp 905	ACA Thr	GTG Val	CCC Pro	TCT Ser	CCC Pro 910	2735
AGG Arg	GAC Asp	CTG Leu	CAG Gln	TTT Phe 915	GTG Val	GAA G1u	GTG Val	ACA Thr	GAC Asp 920	GTG Val	AAG Lys	GTC Val	ACC Thr	ATC Ile 925	ATG Met	2783
TGG Trp	ACA Thr	CCG Pro	CCT Pro 930	GAG Glu	AGT Ser	GCA Ala	GTG Val	ACC Thr 935	GGC Gly	TAC Tyr	CGT Arg	GTG Val	GAT Asp 940	GTG Val	ATC Ile	2831
CCC Pro	GTC Val	AAC Asn 945	CTG Leu	CCT Pro	GGC Gly	GAG Glu	CAC His 950	GGG Gly	CAG Gln	AGG Arg	CTG Leu	CCC Pro 955	ATC Ile	AGC Ser	AGG Arg	2879
AAC Asn	ACC Thr 960	TTT Phe	GCA Ala	GAA Glu	GTC Val	ACC Thr 965	GGG Gly	CTG Leu	TCC Ser	CCT Pro	GGG Gly 970	GTC Val	ACC Thr	TAT Tyr	TAC Tyr	2927
TTC Phe 975	Lys	GTC Val	TTT Phe	GCA Ala	GTG Val 980	AGC Ser	CAT His	GGG Gly	AGG Arg	GAG G1u 985	AGC Ser	AAG Lys	CCT Pro	CTG Leu	ACT Thr 990	2975
GCT Ala	CAA Gln	CAG Gln	ACA Thr	ACC Thr 995	AAA Lys	CTG Leu	GAT Asp	GCT Ala	CCC Pro 1000	Thr	AAC Asn	CTC Leu	CAG Gln	TTT Phe 1005	Val	3023
AAT Asn	GAA Glu	ACT Thr	GAT Asp 1010	Ser	ACT Thr	GTC Val	CTG Leu	GTG Val 1015	Arg	TGG Trp	ACT Thr	CCA Pro	CCT Pro 1020	Arg	GCC Ala	3071
CAG Gln	ATA Ile	ACA Thr 1025	GGA Gly	TAC Tyr	CGA Arg	CTG Leu	ACC Thr 1030	Val	GGC Gly	CTT Leu	ACC Thr	CGA Arg 1035	Arg	GGC Gly	CAG Gln	3119
CCC Pro	AGG Arg 1040	Gln	TAC Tyr	AAT Asn	GTG Val	GGT Gly 1045	Pro	TCT Ser	GTC Val	TCC Ser	AAG Lys 1050	Tyr	CCC Pro	CTG- Leu	AGG Arg	3167
AAT Asn 1055	Leu	CAG Gln	CCT Pro	GCA Ala	TCT Ser 1060	Glu	TAC Tyr	ACC Thr	GTA Val	TCC Ser 1065	Leu	GTG Val	GCC Ala	ATA Ile	AAG Lys 1070	3215

GGC Gly	AAC Asn	CAA Gln	GAG Glu	AGC Ser 107	Pro	AAA Lys	GCC Ala	ACT Thr	GGA Gly 108	Val	TTT Phe	ACC Thr	ACA Thr	CTG Leu 108	Gln	3263
CCT Pro	GGG Gly	AGC Ser	TCT Ser 109	Ile	CCA Pro	CCT Pro	TAC Tyr	AAC Asn 109	Thr	GAG Glu	GTG Val	ACT Thr	GAG Glu 110	Thr	ACC Thr	3311
ATC Ile	GTG Val	ATC Ile 110	Thr	TGG Trp	ACG Thr	CCT Pro	GCT Ala 111	Pro	AGA Arg	ATT Ile	GGT Gly	TTT Phe 111	Lys	CTG Leu	GGT Gly	3359
GTA Val	CGA Arg 1120	Pro	AGC Ser	CAG Gln	GGA Gly	GGA Gly 1125	Glu	GCA Ala	CCA Pro	CGA Arg	GAA Glu 1130	Val	ACT Thr	TCA Ser	GAC Asp	3407
TCA Ser 113	Gly	AGC Ser	ATC Ile	GTT Val	GTG Val 114	TCC Ser O	GGC Gly	TTG Leu	ACT Thr	CCA Pro 114	Gly	GTA Val	GAA Glu	TAC Tyr	GTC Val 1150	3455
TAC Tyr	ACC Thr	ATC Ile	CAA Gln	GTC Val 115	Leu	AGA Arg	GAT Asp	GGA Gly	CAG Gln 1160	Glu	AGA Arg	GAT Asp	GCG Ala	CCA Pro 116	Ile	3503
GTA Val	AAC Asn	AAA Lys	GTG Val 1170	Val	ACA Thr	CCA Pro	TTG Leu	TCT Ser 117	Pro	CCA Pro	ACA Thr	AAC Asn	TTG Leu 1180	His	CTG Leu	3551
GAG Glu	GCA Ala	AAC Asn 118	Pro	GAC Asp	ACT Thr	GGA Gly	GTG Val 1190	Leu	ACA Thr	GTC Val	TCC Ser	TGG Trp 1199	Glu	AGG Arg	AGC Ser	3599
ACC Thr	ACC Thr 1200	Pro	GAC Asp	ATT Ile	ACT Thr	GGT Gly 1205	Tyr	AGA Arg	ATT Ile	ACC Thr	ACA Thr 1210	Thr	CCT Pro	ACA Thr	AAC Asn	3647
GGC Gly 121	G1n	CAG Gln	GGA Gly	AAT Asn	TCT Ser 1220	TTG Leu)	GAA Glu	GAA Glu	GTG Val	GTC Val 1225	His	GCT Ala	GAT Asp	CAG Gln	AGC Ser 1230	3695
TCC Ser	TGC Cys	ACT Thr	TTT Phe	GAT Asp 1235	Asn	CTG Leu	AGT Ser	CCC Pro	GGC Gly 1240	Leu	GAG Glu	TAC Tyr	AAT Asn	GTC Val 1245	Ser	3743
GTT Val	TAC Tyr	ACT Thr	GTC Val 1250	Lys	GAT Asp	GAC Asp	AAG Lys	GAA Glu 1255	Ser	GTC Val	CCT Pro	ATC Ile	TCT Ser 1260	Asp	ACC Thr	3791
ATC Ile	He	CCA Pro 1265	Glu	GTG Val	CCC Pro	CAA Gln	CTC Leu 1270	Thr	GAC Asp	CTA Leu	Ser	TTT Phe 1275	Val	GAT Asp	ATA Ile	3839
ACC Thr	GAT Asp 1280	Ser	AGC Ser	ATC Ile	GGC Gly	CTG Leu 1285	Arg	TGG Trp	ACC Thr	CCG Pro	CTA Leu 1290	Asn	TCT Ser	TCC Ser	ACC Thr	3887

	Пe					ACA Thr					Gly					3935
					Val	TAC Tyr				Gly					Thr	3983
				Gly		GAC Asp			Ile					Leu		4031
			Glu			CCT Pro		Thr					Thr			4079
		Pro				CGA Arg 1365	Phe					Pro				4127
	Val					CCC Pro)					Leu					4175
GTG Val	CGT Arg	TAC Tyr	TCA Ser	CCT Pro 1395	Val	AAA Lys	AAT Asn	GAG Glu	GAA Glu 1400	Asp	GTT Val	GCA Ala	GAG Glu	TTG Leu 1405	Ser	4223
				Asp		GCA Ala			Leu					Pro		4271
			Val			GTC Val		Ser					His			4319
		Leu				CAG Gln 1445	Lys					Ser				4367
	Asp					ACT Thr					Thr					4415
					He	ACT Thr				Пe					Glu	4463
				Arg		CGA Arg			Arg					Arg		4511

TCC Ser	ATC Ile	ACC Thr 150	Leu	ACC Thr	AAC Asn	CTC Leu	ACT Thr 1510	Pro	GGC Gly	ACA Thr	GAG Glu	TAT Tyr 151	Val	GTC Val	AGC Ser	45	59
ATC Ile	GTT Val 152	Ala	CTT Leu	AAT Asn	GGC Gly	AGA Arg 152	Glu	GAA Glu	AGT Ser	CCC Pro	TTA Leu 153	Leu	ATT Ile	GGC Gly	CAA Gln	46	07
CAA Gln 153	TCA Ser 5	ACA Thr	GTT Val	TCT Ser	GAT Asp 1540	Val	CCG Pro	AGG Arg	GAC Asp	CTG Leu 154	Glu	GTT Val	GTT Val	GCT Ala	GCG Ala 1550	46	55
ACC Thr	CCC Pro	ACC Thr	AGC Ser	CTA Leu 155	Leu	ATC Ile	AGC Ser	TGG Trp	GAT Asp 1560	Ala	CCT Pro	GCT Ala	GTC Val	ACA Thr 156	Val	47(03
AGA Arg	TAT Tyr	TAC Tyr	AGG Arg 1570	Ile	ACT Thr	TAC Tyr	GGA Gly	GAA Glu 157	Thr	GGA Gly	GGA Gly	AAT Asn	AGC Ser 1580	Pro	GTC Val	47!	51
CAG Gln	GAG Glu	TTC Phe 158!	Thr	GTG Val	CCT Pro	GGG Gly	AGC Ser 1590	Lys	TCT Ser	ACA Thr	GCT Ala	ACC Thr 159	Ile	AGC Şer	GGC Gly	479	99
CTT Leu	AAA Lys 1600	Pro	GGA Gly	GTT Val	GAT Asp	TAT Tyr 1605	Thr	ATC Ile	ACT Thr	GTG Val	TAT Tyr 1610	Ala	GTC Val	ACT Thr	GGC Gly	484	47
CGT Arg 161	GGA Gly 5	GAC Asp	AGC Ser	CCC Pro	GCA Ala 1620	Ser	AGC Ser	AAG Lys	CCA Pro	ATT Ile 1625	Ser	ATT Ile	AAT Asn	TAC Tyr	CGA Arg 1630	489	95
ACA Thr	GAA Glu	ATT Ile	GAC Asp	AAA Lys 1635	Pro	TCC Ser	CAG Gln	ATG Met	CAA Gln 1640	Val	ACC Thr	GAT Asp	GTT Val	CAG Gln 1645	Asp	494	43
AAC Asn	AGC Ser	ATT Ile	AGT Ser 1650	Val	AAG Lys	TGG Trp	CTG Leu	CCT Pro 1655	Ser	AGT Ser	TCC Ser	CCT Pro	GTT Val 1660	Thr	GGT Gly	499	91
TAC Tyr	AGA Arg	GTA Val 1665	Thr	ACC Thr	ACT Thr	CCC Pro	AAA Lys 1670	Asn	GGA Gly	CCA Pro	GGA Gly	CCA Pro 1675	Thr	AAA Lys	ACT Thr	503	39
	ACT Thr 1680	Ala					Thr					G1u				508	37
CCC Pro 1695	ACA Thr	GTG Val	GAG Glu	TAT Tyr	GTG Val 1700	Val	AGT Ser	GTC Val	TAT Tyr	GCT Ala 1705	Gln	AAT Asn	CCA Pro	AGC Ser	GGA Gly 1710	513	35
GAG Glu	AGT Ser	CAG Gln	CCT Pro	CTG Leu 1715	Val	CAG Gln	ACT Thr	GCA Ala	GTA Val 1720	Thr	AAC Asn	ATT Ile	GAT Asp	CGC Arg 1725	Pro	518	33

AAA Lys	GGA Gly	CTG Leu	GCA Ala 173	Phe	ACT Thr	GAT Asp	GTG Val	GAT Asp 173	Val	GAT Asp	TCC Ser	ATC Ile	AAA Lys 1740	Ile	GCT Ala	5231
TGG Trp	GAA Glu	AGC Ser 174	Pro	CAG Gln	GGG Gly	CAA Gln	GTT Val 1750	Ser	AGG Arg	TAC Tyr	AGG Arg	GTG Val 175	Thr	TAC Tyr	TCG Ser	5279
	CCT Pro 176	Glu					Glu					Pro				5327
GAA Glu 177	GAC Asp 5	ACT Thr	GCA Ala	GAG Glu	CTG Leu 1780	Gln	GGC Gly	CTC Leu	AGA Arg	CCG Pro 178	Gly	TCT Ser	GAG Glu	TAC Tyr	ACA Thr 1790	5375
GTC Val	AGT Ser	GTG Val	GTT Val	GCC Ala 179	Leu	CAC His	GAT Asp	GAT Asp	ATG Met 1800	Glu	AGC Ser	CAG Gln	CCC Pro	CTG Leu 180!	Ile	5423
GGA Gly	ACC Thr	CAG Gln	TCC Ser 1810	Thr	GCT Ala	ATT Ile	CCT Pro	GCA Ala 1815	Pro	ACT Thr	GAC Asp	CTG Leu	AAG Lys 1820	Phe	ACT Thr	5471
CAG Gln	GTC Val	ACA Thr 182	Pro	ACA Thr	AGC Ser	CTG Leu	AGC Ser 1830	Ala	CAG Gln	TGG Trp	ACA Thr	CCA Pro 1835	Pro	AAT Asn	GTT Val	5519
CAG Gln	CTC Leu 1840	Thr	GGA Gly	TAT Tyr	CGA Arg	GTG Val 1845	Arg	GTG Val	ACC Thr	CCC Pro	AAG Lys 1850	Glu	AAG Lys	ACC Thr	GGA Gly	5567
CCA Pro 185	ATG Met 5	AAA Lys	GAA Glu	ATC Ile	AAC Asn 1860	Leu	GCT Ala	CCT Pro	GAC Asp	AGC Ser 1865	Ser	TCC Ser	GTG Val	GTT Val	GTA Val 1870	5615
TCA Ser	GGA Gly	CTT Leu	ATG Met	GTG Val 1875	Ala	ACC Thr	AAA Lys	TAT Tyr	GAA Glu 1880	Val	AGT Ser	GTC Val	TAT Tyr	GCT Ala 1885	Leu	5663
AAG Lys	GAC Asp	ACT Thr	TTG Leu 1890	Thr	AGC Ser	AGA Arg	CCA Pro	GCT Ala 1895	Gln	GGT Gly	GTT Val	GTC Val	ACC Thr 1900	Thr	CTG Leu	5711
	AAT Asn		Ser					Ala					Ala			5759
	ACC Thr 1920	He					Arg					Thr				5807

TTC Phe 193	Gln	GTT Val	GAT Asp	GCC Ala	GTT Val 194	Pro	GCC Ala	AAT Asn	GGC Gly	CAG Gln 194	Thr	CCA Pro	ATC Ile	CAG Gln	AGA Arg 1950	5855
ACC Thr	ATC Ile	AAG Lys	CCA Pro	GAT Asp 195	Val	AGA Arg	AGC Ser	TAC Tyr	ACC Thr 1 9 6	Ile	ACA Thr	GGT Gly	TTA Leu	CAA Gln 196	Pro	5903
GGC Gly	ACT Thr	GAC Asp	TAC Tyr 197	AAG Lys O	ATC Ile	TAC Tyr	CTG Leu	TAC Tyr 197	Thr	TTG Leu	AAT Asn	GAC Asp	AAT Asn 198	Ala	CGG Arg	5951
AGC Ser	TCC Ser	CCT Pro 198	۷a٦	GTC Val	ATC Ile	GAC Asp	GCC Ala 1990	Ser	ACT Thr	GCC Ala	ATT Ile	GAT Asp 199	Ala	CCA Pro	TCC Ser	5999
AAC Asn	CTG Leu 200	Arg	TTC Phe	CTG Leu	GCC Ala	ACC Thr 200	Thr	CCC Pro	AAT Asn	TCC Ser	TTG Leu 201	Leu	GTA Val	TCA Ser	TGG Trp	6047
CAG Gln 201	Pro	CCA Pro	CGT Arg	GCC Ala	AGG Arg 2020	Пe	ACC Thr	GGC Gly	TAC Tyr	ATC Ile 202	Пe	AAG Lys	TAT Tyr	GAG Glu	AAG Lys 2030	6095
CCT Pro	GGG Gly	TCT Ser	CCT Pro	CCC Pro 203	Arg	GAA Glu	GTG Val	GTC Val	CCT Pro 2040	Arg	CCC Pro	CGC Arg	CCT Pro	GGT G1 y 204!	Val	6143
ACA Thr	GAG Glu	GCT Ala	ACT Thr 2050	ATT Ile O	ACT Thr	GGC Gly	CTG Leu	GAA Glu 2055	Pro	GGA Gly	ACC Thr	GAA Glu	TAT Tyr 2060	Thr	ATT Ile	6191
TAT Tyr	GTC Val	ATT Ile 206	Ala	CTG Leu	AAG Lys	AAT Asn	AAT Asn 2070	Gln	AAG Lys	AGC Ser	GAG Glu	CCC Pro 2075	Leu	ATT Ile	GGA Gly	6239
AGG Arg	AAA Lys 2080	Lys	ACA Thr	GAC Asp	GAG G1 u	CTT Leu 2085	Pro	CAA Gln	CTG Leu	GTA Val	ACC Thr 2090	Leu	CCA Pro	CAC His	CCC Pro	6287
AAT Asn 2095	Leu	CAT His	GGA Gly	CCA Pro	GAG Glu 2100	He	TTG Leu	GAT Asp	GTT Val	CCT Pro 2105	Ser	ACA Thr	GTT Val	CAA Gln	AAG Lys 2110	6335
ACC Thr	CCT Pro	TTC Phe	GTC Val	ACC Thr 2115	His	CCT Pro	GGG Gly	TAT Tyr	GAC Asp 2120	Thr	GGA Gly	AAT Asn	GGT Gly	ATT Ile 2125	Gln	6383
CTT Leu	CCT Pro	GGC Gly	ACT Thr 2130	TCT Ser	GGT Gly	CAG Gln	Gln	CCC Pro 2135	Ser	GTT Val	GGG Gly	CAA Gln	CAA Gln 2140	Met	ATC Ile	6431
TTT Phe	GAG Glu	GAA Glu 2145	His	GGT Gly	TTT Phe	Arg	CGG Arg 2150	Thr	ACA Thr	CCG Pro	CCC Pro	ACA Thr 2155	Thr	GCC Ala	ACC Thr	6479

CCC ATA AGG Pro Ile Arg 2160	CAT AGG CCA His Arg Pro	AGA CCA Arg Pro 2165	TAC CCG Tyr Pro	CCG AAT Pro Asn 217	Val Gly	CAA G Gln G	AA 6527 Iu
GCT CTC TCT Ala Leu Ser 2175	CAG ACA ACC Gln Thr Thr 218	Ile Ser	TGG GCC Trp Ala	CCA TTC Pro Phe 2185	CAG GAC Gln Asp	Thr S	CT 6575 er 190
GAG TAC ATC . Glu Tyr Ile	ATT TCA TGT Ile Ser Cys 2195	CAT CCT His Pro	GTT GGC Val Gly 2200	Thr Asp	GAA GAA Glu Glu	CCC T Pro Le 2205	TA 6623 eu
CAG TTC AGG Gln Phe Arg	GTT CCT GGA Val Pro Gly 2210	Thr Ser	ACC AGT Thr Ser 2215	GCC ACT Ala Thr	CTG ACA Leu Thr 2220	Gly Le	C 6671 eu
ACC AGA GGT Thr Arg Gly 2225	Ala Thr Tyr	AAC ATC A Asn Ile 2230	Ile Val	GAG GCA Glu Ala	CTG AAA Leu Lys 2235	GAC CA	AG 6719 n
CAG AGG CAT A Gln Arg His 2240	AAG GTT CGG Lys Val Arg	GAA GAG Glu Glu 2245	GTT GTT Val Val	ACC GTG Thr Val 2250	Gly Asn	TCT GT Ser Va	C 6767
AAC GAA GGC Asn Glu Gly 2255	TTG AAC CAA Leu Asn Gln 2260	Pro Thr	GAT GAC Asp Asp	TCG TGC Ser Cys 2265	TTT GAC Phe Asp	Pro Ty	AC 6815 270
ACA GTT TCC (Thr Val Ser I	CAT TAT GCC His Tyr Ala 2275	GTT GGA (Val Gly	GAT GAG Asp Glu 2280	Trp Glu	CGA ATG Arg Met	TCT GA Ser G1 2285	AA 6863 u
TCA GGC TTT / Ser Gly Phe I	AAA CTG TTG Lys Leu Leu 2290	Cys Gln (TGC TTA Cys Leu 2295	GGC TTT Gly Phe	GGA AGT Gly Ser 2300	Gly Hi	T 6911
TTC AGA TGT (Phe Arg Cys / 2305	Asp Ser Ser		Cys His				
AAG ATT GGA (Lys Ile Gly (2320					Gly Gln		
AGC TGC ACA Ser Cys Thr (2335	TGT CTT GGG Cys Leu Gly 2340	Asn Gly I	Lys Gly	GAA TTC Glu Phe 2345	AAG TGT Lys Cys	Asp Pr	T 7055 o 50
CAT GAG GCA A His Glu Ala	ACG TGT TAC Thr Cys Tyr 2355	GAT GAT (Asp Asp (GGG AAG Gly Lys 2360	Thr Tyr	CAC GTA His Val	GGA GA Gly Gl 2365	A 7103 u

CAG Gln	TGG Trp	CAG Gln	AAG Lys 2370	Glu	TAT Tyr	CTC Leu	GGT Gly	GCC Ala 237	Ile	TGC Cys	TCC Ser	TGC Cys	ACA Thr 238	TGC Cys O	TTT Phe		7151
GGA Gly	GGC Gly	CAG G1n 238	Arg	GGC Gly	TGG Trp	CGC Arg	TGT Cys 2390	Asp	AAC Asn	TGC Cys	CGC Arg	AGA Arg 239	Pro	GGG Gly	GGT Gly		7199
GAA Glu	CCC Pro 2400	Ser	CCC Pro	GAA Glu	GGC Gly	ACT Thr 2405	Thr	GGC Gly	CAG Gln	TCC Ser	TAC Tyr 2410	Asn	CAG Gln	TAT Tyr	TCT Ser		7247
CAG Gln 2415	Arg	TAC Tyr	CAT His	CAG Gln	AGA Arg 2420	Thr	AAC Asn	ACT Thr	AAT Asn	GTT Val 2425	Asn	TGC Cys	CCA Pro	ATT Ile	GAG Glu 2430		7295
TGC Cys	TTC Phe	ATG Met	CCT Pro	TTA Leu 2435	Asp	GTA Val	CAG G1n	GCT Ala	GAC Asp 2440	Arg	GAA Glu	GAT Asp	TCC Ser	CGA Arg 2445	Glu		7343
TAAA	TCAT	TCT T	TTCC#	AATCO	A GA	AGGAA	CAAG	CAT	rgtct	стс	TGCC	CAAGA	ATC (CATCT	TAAACT	ſ	7403
GGAG	TGAT	GT 1	ragc <i>a</i>	AGACO	C AG	CTTA	GAGT	TCT	тстт	тст	TTCT	TAAG	scc (CTTTG	стсто	à	7463
GAGG	AAGT	TC 7	CCAG	CTTC	A GC	TCAA	CTCA	CAG	CTTC	TCC	AAGO	ATCA	ACC (CTGGG	AGTT	Γ	7523
CCTG	AGGG	I TT	тсто	CATAA	A TO	AGG	CTGC	ACA	TTGC	CTG	ттст	GCTT	CG A	AAGTA	TTCA	4	7583
TACC	GCTC	AG 7	TATT	ГТААА	T GA	AGTG	ATTO	TAA	GATT	TGG	TTTG	GGAT	CA A	ATAGG	AAAGC		7643
ATAT	GCAG	icc A	ACCA	AGAT	G CA	AATG	TTTT	GAA	ATGA	TAT	GACC	AAAA	ר דדו	TAAG	TAGGA	١	7703
AAGT	CACC	CA A	ACAC	ттст	G CT	TTCA	CTTA	AGT	GTCT	GGC	CCGC	AATA	CT (STAGG	AACAA	4	7763
GCAT	GATC	TT G	TTAC	TGTG	A TA	TTTT	TAAA	ATC	CACA	GTA							7803

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2446 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Leu Arg Gly Pro Gly Pro Gly Leu Leu Leu Ala Val Leu Cys

Leu Gly Thr Ala Val Pro Ser Thr Gly Ala Ser Lys Ser Lys Arg Gln 20 25 30

Ala Gln Gln Met Val Gln Pro Gln Ser Pro Val Ala Val Ser Gln Ser 40 Lys Pro Gly Cys Tyr Asp Asn Gly Lys His Tyr Gln Ile Asn Gln Gln Trp Glu Arg Thr Tyr Leu Gly Asn Val Leu Val Cys Thr Cys Tyr Gly Gly Ser Arg Gly Phe Asn Cys Glu Ser Lys Pro Glu Ala Glu Glu Thr Cys Phe Asp Lys Tyr Thr Gly Asn Thr Tyr Arg Val Gly Asp Thr Tyr Glu Arg Pro Lys Asp Ser Met Ile Trp Asp Cys Thr Cys Ile Gly Ala 120. Gly Arg Gly Arg Ile Ser Cys Thr Ile Ala Asn Arg Cys His Glu Gly Gly Gln Ser Tyr Lys Ile Gly Asp Thr Trp Arg Arg Pro His Glu Thr Gly Gly Tyr Met Leu Glu Cys Val Cys Leu Gly Asn Gly Lys Gly Glu Trp Thr Cys Lys Pro Ile Ala Glu Lys Cys Phe Asp His Ala Ala Gly Thr Ser Tyr Val Val Gly Glu Thr Trp Glu Lys Pro Tyr Gln Gly Trp 200 Met Met Val Asp Cys Thr Cys Leu Gly Glu Gly Ser Gly Arg Ile Thr Cys Thr Ser Arg Asn Arg Cys Asn Asp Gln Asp Thr Arg Thr Ser Tyr 235 Arg Ile Gly Asp Thr Trp Ser Lys Lys Asp Asn Arg Gly Asn Leu Leu Gln Cys Ile Cys Thr Gly Asn Gly Arg Gly Glu Trp Lys Cys Glu Arg 265 His Thr Ser Val Gln Thr Thr Ser Ser Gly Ser Gly Pro Phe Thr Asp Val Arg Ala Ala Val Tyr Gln Pro Gln Pro His Pro Gln Pro Pro 295

Tyr Gly His Cys Val Thr Asp Ser Gly Val Val Tyr Ser Val Gly Met

315

Gln Trp Leu Lys Thr Gln Gly Asn Lys Gln Met Leu Cys Thr Cys Leu Gly Asn Gly Val Ser Cys Gln Glu Thr Ala Val Thr Gln Thr Tyr Gly 345 Gly Asn Leu Asn Gly Glu Pro Cys Val Leu Pro Phe Thr Tyr Asn Gly Arg Thr Phe Tyr Ser Cys Thr Thr Glu Gly Arg Gln Asp Gly His Leu 375 380 Trp Cys Ser Thr Thr Ser Asn Tyr Glu Gln Asp Gln Lys Tyr Ser Phe 390 400 Cys Thr Asp His Thr Val Leu Val Gln Thr Gln Gly Gly Asn Ser Asn 410 Gly Ala Leu Cys His Phe Pro Phe Leu Tyr Asn Asn His Asn Tyr Thr 420 425 Asp Cys Thr Ser Glu Gly Arg Arg Asp Asn Met Lys Trp Cys Gly Thr Thr Gln Asn Tyr Asp Ala Asp Gln Lys Phe Gly Phe Cys Pro Met Ala 455 Ala His Glu Glu Ile Cys Thr Thr Asn Glu Gly Val Met Tyr Arg Ile Gly Asp Gln Trp Asp Lys Gln His Asp Met Gly His Met Met Arg Cys 490 Thr Cys Val Gly Asn Gly Arg Gly Glu Trp Thr Cys Ile Ala Tyr Ser 505 Gln Leu Arg Asp Gln Cys Ile Val Asp Asp Ile Thr Tyr Asn Val Asn 520 Asp Thr Phe His Lys Arg His Glu Glu Gly His Met Leu Asn Cys Thr 530 Cys Phe Gly Gln Gly Arg Gly Arg Trp Lys Cys Asp Pro Val Asp Gln Cys Gln Asp Ser Glu Thr Gly Thr Phe Tyr Gln Ile Gly Asp Ser Trp 565 Glu Lys Tyr Val His Gly Val Arg Tyr Gln Cys Tyr Cys Tyr Gly Arg 585 Gly Ile Gly Glu Trp His Cys Gln Pro Leu Gln Thr Tyr Pro Ser Ser 595 600 605

Ser Gly Pro Val Glu Val Phe Ile Thr Glu Thr Pro Ser Gln Pro Asn 615 Ser His Pro Ile Gln Trp Asn Ala Pro Gln Pro Ser His Ile Ser Lys 630 Tyr Ile Leu Arg Trp Arg Pro Lys Asn Ser Val Gly Arg Trp Lys Glu Ala Thr Ile Pro Gly His Leu Asn Ser Tyr Thr Ile Lys Gly Leu Lys 665 Pro Gly Val Val Tyr Glu Gly Gln Leu Ile Ser Ile Gln Gln Tyr Gly His Gln Glu Val Thr Arg Phe Asp Phe Thr Thr Thr Ser Thr Ser Thr Pro Val Thr Ser Asn Thr Val Thr Gly Glu Thr Thr Pro Phe Ser Pro 710 Leu Val Ala Thr Ser Glu Ser Val Thr Glu Ile Thr Ala Ser Ser Phe 730 Val Val Ser Trp Val Ser Ala Ser Asp Thr Val Ser Gly Phe Arg Val Glu Tyr Glu Leu Ser Glu Glu Gly Asp Glu Pro Gln Tyr Leu Asp Leu Pro Ser Thr Ala Thr Ser Val Asn Ile Pro Asp Leu Leu Pro Gly Arg Lys Tyr Ile Val Asn Val Tyr Gln Ile Ser Glu Asp Gly Glu Gln Ser 790 795 Leu Ile Leu Ser Thr Ser Gln Thr Thr Ala Pro Asp Ala Pro Pro Asp 810 Pro Thr Val Asp Gln Val Asp Asp Thr Ser Ile Val Val Arg Trp Ser 820 825 Arg Pro Gln Ala Pro Ile Thr Gly Tyr Arg Ile Val Tyr Ser Pro Ser 840 Val Glu Gly Ser Ser Thr Glu Leu Asn Leu Pro Glu Thr Ala Asn Ser 850 860 Val Thr Leu Ser Asp Leu Gln Pro Gly Val Gln Tyr Asn Ile Thr Ile Tyr Ala Val Glu Glu Asn Gln Glu Ser Thr Pro Val Val Ile Gln Gln

- Glu Thr Thr Gly Thr Pro Arg Ser Asp Thr Val Pro Ser Pro Arg Asp 900 905 910
- Leu Gln Phe Val Glu Val Thr Asp Val Lys Val Thr Ile Met Trp Thr 915 920 925
- Pro Pro Glu Ser Ala Val Thr Gly Tyr Arg Val Asp Val Ile Pro Val 930 935 940
- Asn Leu Pro Gly Glu His Gly Gln Arg Leu Pro Ile Ser Arg Asn Thr 945 950 955 960
- Phe Ala Glu Val Thr Gly Leu Ser Pro Gly Val Thr Tyr Tyr Phe Lys 965 970 975
- Val Phe Ala Val Ser His Gly Arg Glu Ser Lys Pro Leu Thr Ala Gln 980 985 990
- Gln Thr Thr Lys Leu Asp Ala Pro Thr Asn Leu Gln Phe Val Asn Glu 995 1000 1005
- Thr Asp Ser Thr Val Leu Val Arg Trp Thr Pro Pro Arg Ala Gln Ile 1010 1015 1020
- Thr Gly Tyr Arg Leu Thr Val Gly Leu Thr Arg Arg Gly Gln Pro Arg 1025 1030 1035 1040
- Gln Tyr Asn Val Gly Pro Ser Val Ser Lys Tyr Pro Leu Arg Asn Leu 1045 1050 1055
- Gln Pro Ala Ser Glu Tyr Thr Val Ser Leu Val Ala Ile Lys Gly Asn 1060 1065 1070
- Gln Glu Ser Pro Lys Ala Thr Gly Val Phe Thr Thr Leu Gln Pro Gly 1075 1080 1085
- Ser Ser Ile Pro Pro Tyr Asn Thr Glu Val Thr Glu Thr Thr Ile Val 1090 1095 1100
- Ile Thr Trp Thr Pro Ala Pro Arg Ile Gly Phe Lys Leu Gly Val Arg 1105 1110 1115 1120
- Pro Ser Gln Gly Glu Ala Pro Arg Glu Val Thr Ser Asp Ser Gly 1125 1130 1135
- Ser Ile Val Val Ser Gly Leu Thr Pro Gly Val Glu Tyr Val Tyr Thr 1140 1145 1150
- Ile Gln Val Leu Arg Asp Gly Gln Glu Arg Asp Ala Pro Ile Val Asn 1155 1160 1165
- Lys Val Val Thr Pro Leu Ser Pro Pro Thr Asn Leu His Leu Glu Ala 1170 1180

- Asn Pro Asp Thr Gly Val Leu Thr Val Ser Trp Glu Arg Ser Thr Thr 1185 1190 1195 1200
- Pro Asp Ile Thr Gly Tyr Arg Ile Thr Thr Thr Pro Thr Asn Gly Gln 1205 1210 1215
- Gln Gly Asn Ser Leu Glu Glu Val Val His Ala Asp Gln Ser Ser Cys 1220 1225 1230
- Thr Phe Asp Asn Leu Ser Pro Gly Leu Glu Tyr Asn Val Ser Val Tyr 1235 1240 1245
- Thr Val Lys Asp Asp Lys Glu Ser Val Pro Ile Ser Asp Thr Ile Ile 1250 1255 1260
- Pro Glu Val Pro Gln Leu Thr Asp Leu Ser Phe Val Asp Ile Thr Asp 1265 1270 1275 1280
- Ser Ser Ile Gly Leu Arg Trp Thr Pro Leu Asn Ser Ser Thr Ile Ile 1285 1290 1295
- Gly Tyr Arg Ile Thr Val Val Ala Ala Gly Glu Gly Ile Pro Ile Phe 1300 1305 1310
- Glu Asp Phe Val Tyr Ser Ser Val Gly Tyr Tyr Thr Val Thr Gly Leu 1315 1320 1325
- Glu Pro Gly Ile Asp Tyr Asp Ile Ser Val Ile Thr Leu Ile Asn Gly 1330 1340
- Gly Glu Ser Ala Pro Thr Thr Leu Thr Gln Gln Thr Ala Val Pro Pro 1345 1350 1360
- Pro Thr Asp Leu Arg Phe Thr Asn Ile Gly Pro Asp Thr Met Arg Val 1365 1370 1375
- Thr Trp Ala Pro Pro Pro Ser Ile Asp Leu Thr Asn Phe Leu Val Arg 1380 1385 1390
- Tyr Ser Pro Val Lys Asn Glu Glu Asp Val Ala Glu Leu Ser Ile Ser 1395 1400 1405
- Pro Ser Asp Asn Ala Val Val Leu Thr Asn Leu Leu Pro Gly Thr Glu 1410 1415 1420
- Tyr Val Val Ser Val Ser Ser Val Tyr Glu Gln His Glu Ser Thr Pro 1425 1430 1435 1440
- Leu Arg Gly Arg Gln Lys Thr Gly Leu Asp Ser Pro Thr Gly Ile Asp 1445 1450 1455
- Phe Ser Asp Ile Thr Ala Asn Ser Phe Thr Val His Trp Ile Ala Pro 1460 1465 1470

- Arg Ala Thr Ile Thr Gly Tyr Arg Ile Arg His His Pro Glu His Phe 1475 1480 1485
- Ser Gly Arg Pro Arg Glu Asp Arg Val Pro His Ser Arg Asn Ser Ile 1490 1495 1500
- Thr Leu Thr Asn Leu Thr Pro Gly Thr Glu Tyr Val Val Ser Ile Val 1505 1510 1515 1520
- Ala Leu Asn Gly Arg Glu Glu Ser Pro Leu Leu Ile Gly Gln Gln Ser 1525 1530 1535
- Thr Val Ser Asp Val Pro Arg Asp Leu Glu Val Val Ala Ala Thr Pro 1540 1545 1550
- Thr Ser Leu Leu Ile Ser Trp Asp Ala Pro Ala Val Thr Val Arg Tyr 1555 1560 1565
- Tyr Arg Ile Thr Tyr Gly Glu Thr Gly Gly Asn Ser Pro Val Gln Glu 1570 1580
- Phe Thr Val Pro Gly Ser Lys Ser Thr Ala Thr Ile Ser Gly Leu Lys 1585 1590 1595 1600
- Pro Gly Val Asp Tyr Thr Ile Thr Val Tyr Ala Val Thr Gly Arg Gly 1605 1610 1615
- Asp Ser Pro Ala Ser Ser Lys Pro Ile Ser Ile Asn Tyr Arg Thr Glu 1620 1625 1630
- Ile Asp Lys Pro Ser Gln Met Gln Val Thr Asp Val Gln Asp Asn Ser 1635 1640 1645
- Ile Ser Val Lys Trp Leu Pro Ser Ser Ser Pro Val Thr Gly Tyr Arg 1650 1655 1660
- Val Thr Thr Pro Lys Asn Gly Pro Gly Pro Thr Lys Thr 1665 1670 1675 1680
- Ala Gly Pro Asp Gln Thr Glu Met Thr Ile Glu Gly Leu Gln Pro Thr 1685 1690 1695
- Val Glu Tyr Val Val Ser Val Tyr Ala Gln Asn Pro Ser Gly Glu Ser 1700 1705 1710
- Gln Pro Leu Val Gln Thr Ala Val Thr Asn Ile Asp Arg Pro Lys Gly
 1715 1720 1725
- Leu Ala Phe Thr Asp Val Asp Val Asp Ser Ile Lys Ile Ala Trp Glu 1730 1740
- Ser Pro Gln Gly Gln Val Ser Arg Tyr Arg Val Thr Tyr Ser Ser Pro 1745 1750 1755 1760

- Glu Asp Gly Ile His Glu Leu Phe Pro Ala Pro Asp Gly Glu Glu Asp 1765 1770 1775
- Thr Ala Glu Leu Gln Gly Leu Arg Pro Gly Ser Glu Tyr Thr Val Ser 1780 1785 1790
- Val Val Ala Leu His Asp Asp Met Glu Ser Gln Pro Leu Ile Gly Thr 1795 1800 1805
- Gln Ser Thr Ala Ile Pro Ala Pro Thr Asp Leu Lys Phe Thr Gln Val 1810 1815 1820
- Thr Pro Thr Ser Leu Ser Ala Gln Trp Thr Pro Pro Asn Val Gln Leu 1825 1830 1835 1840
- Thr Gly Tyr Arg Val Arg Val Thr Pro Lys Glu Lys Thr Gly Pro Met 1845 1850 1855
- Lys Glu Ile Asn Leu Ala Pro Asp Ser Ser Ser Val Val Val Ser Gly 1860 1865 1870
- Leu Met Val Ala Thr Lys Tyr Glu Val Ser Val Tyr Ala Leu Lys Asp 1875 1880 1885
- Thr Leu Thr Ser Arg Pro Ala Gln Gly Val Val Thr Thr Leu Glu Asn 1890 1895 1900
- Val Ser Pro Pro Arg Arg Ala Arg Val Thr Asp Ala Thr Glu Thr Thr 1905 1910 1915 1920
- Ile Thr Ile Ser Trp Arg Thr Lys Thr Glu Thr Ile Thr Gly Phe Gln
 1925 1930 1935
- Val Asp Ala Val Pro Ala Asn Gly Gln Thr Pro Ile Gln Arg Thr Ile 1940 1945 1950
- Lys Pro Asp Val Arg Ser Tyr Thr Ile Thr Gly Leu Gln Pro Gly Thr 1955 1960 1965
- Asp Tyr Lys Ile Tyr Leu Tyr Thr Leu Asn Asp Asn Ala Arg Ser Ser 1970 1975 1980
- Pro Val Val Ile Asp Ala Ser Thr Ala Ile Asp Ala Pro Ser Asn Leu 1985 1990 1995 2000
- Arg Phe Leu Ala Thr Thr Pro Asn Ser Leu Leu Val Ser Trp Gln Pro 2005 2010 2015
- Pro Arg Ala Arg Ile Thr Gly Tyr Ile Ile Lys Tyr Glu Lys Pro Gly 2020 2030
- Ser Pro Pro Arg Glu Val Val Pro Arg Pro Arg Pro Gly Val Thr Glu 2035 2040 2045

- Ala Thr Ile Thr Gly Leu Glu Pro Gly Thr Glu Tyr Thr Ile Tyr Val 2050 2055 2060
- Ile Ala Leu Lys Asn Asn Gln Lys Ser Glu Pro Leu Ile Gly Arg Lys 2065 2070 2075 2080
- Lys Thr Asp Glu Leu Pro Gln Leu Val Thr Leu Pro His Pro Asn Leu 2085 2090 2095
- His Gly Pro Glu Ile Leu Asp Val Pro Ser Thr Val Gln Lys Thr Pro 2100 2105 2110
- Phe Val Thr His Pro Gly Tyr Asp Thr Gly Asn Gly Ile Gln Leu Pro 2115 2120 2125
- Gly Thr Ser Gly Gln Gln Pro Ser Val Gly Gln Gln Met Ile Phe Glu 2130 2135 2140
- Glu His Gly Phe Arg Arg Thr Thr Pro Pro Thr Thr Ala Thr Pro Ile 2145 2150 2155 2160
- Arg His Arg Pro Arg Pro Tyr Pro Pro Asn Val Gly Gln Glu Ala Leu 2165 2170 2175
- Ser Gln Thr Thr Ile Ser Trp Ala Pro Phe Gln Asp Thr Ser Glu Tyr 2180 2185 2190
- Ile Ile Ser Cys His Pro Val Gly Thr Asp Glu Glu Pro Leu Gln Phe 2195 2200 2205
- Arg Val Pro Gly Thr Ser Thr Ser Ala Thr Leu Thr Gly Leu Thr Arg 2210 2215 2220
- Gly Ala Thr Tyr Asn Ile Ile Val Glu Ala Leu Lys Asp Gln Gln Arg 2225 2230 2235 2240
- His Lys Val Arg Glu Glu Val Val Thr Val Gly Asn Ser Val Asn Glu 2245 2250 2255
- Gly Leu Asn Gln Pro Thr Asp Asp Ser Cys Phe Asp Pro Tyr Thr Val 2260 2265 2270
- Ser His Tyr Ala Val Gly Asp Glu Trp Glu Arg Met Ser Glu Ser Gly 2275 2280 2285
- Phe Lys Leu Leu Cys Gln Cys Leu Gly Phe Gly Ser Gly His Phe Arg 2290 2295 2300
- Cys Asp Ser Ser Arg Trp Cys His Asp Asn Gly Val Asn Tyr Lys Ile 2305 2310 2315 2320
- Gly Glu Lys Trp Asp Arg Gln Gly Glu Asn Gly Gln Met Met Ser Cys 2325 2330 2335

Thr	Cys	Leu	Gly 2340	Asn O	Gly	Lys	Gly	G1u 234!	Phe 5	Lys	Cys	Asp	Pro 235		Glu		
A1 a	Thr	Cys 235!	Tyr 5	Asp	Asp	Gly	Lys 2360	Thr)	Tyr	His	Val	Gly 2365		Gln	Trp	•	.*
Gln	Lys 237	Glu O	Tyr	Leu	Gly	Ala 2375	Ile	Cys	Ser	Cys	Thr 2380		Phe	Gly	Gly		
G1n 238	Arg 5	Gly	Trp	Arg	Cys 2390	Asp)	Asn	Cys	Arg	Arg 239	Pro 5	Gly	Gly	Glu	Pro 2400		
Ser	Pro	Glu	Gly	Thr 2405	Thr	Gly	Gln	Ser	Tyr 2410		Gln	Tyr	Ser	Gln 241			
Tyr	His	Gln	Arg 2420	Thr)	Asn	Thr	Asn	Val 2425	Asn	Cys	Pro	Пe	G1u 2430		Phe		
Met	Pro	Leu 2435	Asp	Val	Gln	Ala	Asp 2440		Glu	Asp	Ser	Arg 2 4 45					
(2)	INFO	ORMAT	ION	FOR	SEQ	ID N	10:3:										
	(i)	(E	N) LE B) TY C) ST	NGTH PE: RAND	: 21 nucl EDNE	TERI 79 b eic SS: line	ase acid sing	pair	`s								
	(ix)) NA	ME/K		CDS 3 1	1962										
		SEQ										٠					
GTC	TAGGA	IGC C	AGCC	CCAC	C CT	TAGA	AAAG								TGC Cys		54
CTA Leu	GTT Val 10	CTA Leu	AGT Ser	GTG Val	GTG Val	GGC Gly 15	ACA Thr	GCA Ala	TGG Trp	ACT Thr	GCA Ala 20	GAT . Asp	AGT Ser	GGT Gly	GAA Glu	1	02
GGT Gly 25	GAC Asp	TTT Phe	CTA Leu	GCT Ala	GAA Glu 30	GGA Gly	GGA Gly	GGC Gly	GTG Val	CGT Arg 35	GGC (Gly	CCA . Pro .	AGG Arg	GTT Val	GTG Val 40	1	50
GAA Glu	AGA Arg	CAT His	CAA Gln	TCT Ser 45	GCC Ala	TGC . Cys	AAA Lys	GAT Asp	TCA Ser 50	GAC Asp	TGG (Trp	CCC Pro	TTC Phe	TGC Cys 55	TCT Ser	1	98

GAT Asp	GAA Glu	GAC Asp	TGG Trp 60	Asn	TAC Tyr	AAA Lys	TGC Cys	CCT Pro 65	Ser	GGC Gly	TGC Cys	AG0 Arg	ATG Met	Lys	GGG	246
TTG Leu	ATT Ile	GAT Asp 75	Glu	GTC Val	AAT Asn	CAA Gln	GAT Asp 80	Phe	ACA Thr	AAC Asn	AGA Arg	ATA Ile 85	Asn	AAG Lys	CTC Leu	294
AAA Lys	AAT Asn 90	Ser	CTA Leu	TTT Phe	GAA Glu	TAT Tyr 95	CAG Gln	AAG Lys	AAC Asn	AAT Asn	AAG Lys 100	Asp	TCT Ser	CAT	TCG Ser	342
TTG Leu 105	lhr	ACT Thr	AAT Asn	ATA Ile	ATG Met 110	Glu	ATT	TTG Leu	AGA Arg	GGC Gly 115	GAT Asp	TTT Phe	TCC Ser	TCA Ser	GCC Ala 120	390
AAT Asn	AAC Asn	CGT Arg	GAT Asp	AAT Asn 125	ACC Thr	TAC Tyr	AAC Asn	CGA Arg	GTG Val 130	TCA Ser	GAG Glu	GAT Asp	CTG Leu	AGA Arg 135	AGC Ser	438
AGA Arg	ATT Ile	GAA Glu	GTC Val 140	CTG Leu	AAG Lys	CGC Arg	AAA Lys	GTC Val 145	ATA Ile	GAA Glu	AAA Lys	GTA Val	CAG Gln 150	CAT His	ATC Ile	486
CAG Gln	CTT Leu	CTG Leu 155	CAG Gln	AAA Lys	AAT Asn	GTT Val	AGA Arg 160	GCT Ala	CAG Gln	TTG Leu	GTT Val	GAT Asp 165	ATG Met	AAA Lys	CGA Arg	534
CTG Leu	GAG Glu 170	GTG Val	GAC Asp	ATT Ile	GAT Asp	ATT Ile 175	AAG Lys	ATC Ile	CGA Arg	TCT Ser	TGT Cys 180	CGA Arg	GGG Gly	TCA Ser	TGG Trp	582
AGT Ser 185	AGG Arg	GCT Ala	TTA Leu	GCT Ala	CGT Arg 190	GAA Glu	GTA Val	GAT Asp	CTG Leu	AAG Lys 195	GAC Asp	TAT Tyr	GAA Glu	GAT Asp	CAG Gln 200	630
GIN	Lys	Gin	CT T Leu	G1u 205	Gln	Val	Ile	Ala	Lys 210	Asp	Leu	Leu	Pro	Ser 215	Arg	678
GAT Asp	AGG Arg	CAA Gln	CAC His 220	TTA Leu	CCA Pro	CTG Leu	ATA Ile	AAA Lys 225	ATG Met	AAA Lys	CCA Pro	GTT Val	CCA Pro 230	GAC Asp	TTG Leu	726
GTT Val	CCC Pro	GGA Gly 235	AAT Asn	TTT Phe	AAG Lys	AGC Ser	CAG Gln 240	CTT Leu	CAG Gln	AAG Lys	GTA Val	CCC Pro 245	CCA Pro	GAG Glu	TGG Trp	774
AAG Lys	GCA Ala 250	TTA Leu	ACA Thr	GAC Asp	ATG Met	CCG Pro 255	CAG Gln	ATG Met	AGA Arg	Met	GAG Glu 260	TTA Leu	GAG Glu	AGA Arg	CCT Pro	822
GGT Gly 265	GGA Gly	AAT Asn	GAG Glu	ATT Ile	ACT Thr 270	CGA Arg	GGA Gly	GGC Gly	Ser	ACC Thr 275	TCT Ser	TAT Tyr	GGA Gly	ACC Thr	GGA G1 y 280	870

TCA Ser	GAG Glu	ACG Thr	GAA Glu	AGC Ser 285	CCC Pro	AGG Arg	AAC Asn	CCT Pro	AGC Ser 290	AGT Ser	GCT Ala	GGA Gly	AGC Ser	TGG Trp 295	AAC Asn	918	ļ
TCT Ser	GGG Gly	AGC Ser	TCT Ser 300	GGA Gly	CCT Pro	GGA Gly	AGT Ser	ACT Thr 305	GGA Gly	AAC Asn	CGA Arg	AAC Asn	CCT Pro 310	GGG Gly	AGC Ser	966	I
TCT Ser	GGG Gly	ACT Thr 315	GGA Gly	GGG Gly	ACT Thr	GCA Ala	ACC Thr 320	TGG Trp	AAA Lys	CCT Pro	GGG Gly	AGC Ser 325	TCT Ser	GGA Gly	CCT Pro	1014	
GGA Gly	AGT Ser 330	GCT Ala	GGA Gly	AGC Ser	TGG Trp	AAC Asn 335	TCT Ser	GGG Gly	AGC Ser	TCT Ser	GGA Gly 340	ACT Thr	GGA Gly	AGT Ser	ACT Thr	1062	
GGA Gly 345	Asn	CAA Gln	AAC Asn	CCT Pro	GGA Gly 350	AGT Ser	CCT Pro	AGA Arg	CCT Pro	GGT Gly 355	AGT Ser	ACC Thr	GGA Gly	ACC Thr	TGG Trp 360	1110	
AAT Asn	CCT Pro	GGC Gly	AGC Ser	TCT Ser 365	GAA Glu	CGC Arg	GGA Gly	AGT Ser	GCT Ala 370	GGG Gly	CAC His	TGG Trp	ACC Thr	TCT Ser 375	GAG Glu	1158	
AGC Ser	TCT Ser	GTA Val	TCT Ser 380	GGT Gly	AGT Ser	ACT Thr	GGA Gly	CAA Gln 385	TGG Trp	CAC His	TCT Ser	GAA G1u	TCT Ser 390	GGA Gly	AGT Ser	1206	
TTT Phe	AGG Arg	CCA Pro 395	GAT Asp	AGC Ser	CCA Pro	GGC Gly	TCT Ser 400	GGG Gly	AAC Asn	GCG Ala	AGG Arg	CCT Pro 405	AAC Asn	AAC Asn	CCA Pro	1254	
GAC Asp	TGG Trp 410	GGC Gly	ACA Thr	TTT Phe	GAA Glu	GAG Glu 415	GTG Val	TCA Ser	GGA Gly	AAT Asn	GTA Val 420	AGT Ser	CCA Pro	GGG Gly	ACA Thr	1302	
AGG Arg 425	AGA Arg	GAG Glu	TAC Tyr	CAC His	ACA Thr 430	GAA Glu	AAA Lys	CTG Leu	GTC Val	ACT Thr 435	AAA Lys	GGA Gly	GAT Asp	AAA Lys	GAG Glu 440	1350	
CTC Leu	AGG Arg	ACT Thr	GGT Gly	AAA Lys 445	GAG Glu	AAG Lys	GTC Val	ACC Thr	TCT Ser 450	GGT Gly	AGC Ser	ACA Thr	ACC Thr	ACC Thr 455	ACG Thr	1398	
CGT Arg	CGT Arg	TCA Ser	TGC Cys 460	TCT Ser	AAA Lys	ACC Thr	GTT Val	ACT Thr 465	AAG Lys	ACT Thr	GTT Val	ATT Ile	GGT Gly 470	CCT Pro	GAT [*] Asp	1446	
GGT Gly	CAC His	AAA Lys 475	GAA Glu	GTT Val	ACC Thr	Lys	GAA Glu 480	GTG Val	GTG Val	ACC Thr	Ser	GAA G1u 485	GAT Asp	GGT Gly	TCT Ser	1494	

GAC Asp	TGT Cys 490	CCC Pro	GAG Glu	GCA Ala	ATG Met	GAT Asp 495	TTA Leu	GGC Gly	ACA Thr	TTG Leu	TCT- Ser 50 0	GGC Gly	ATA Ile	GGT Gly	ACT Thr	1542
CTG Leu 505	GAT Asp	GGG Gly	TTC Phe	CGT Arg	CAT His 510	AGG Arg	CAC His	CCT Pro	GAT Asp	GAA Glu 515	GCT Ala	GCC Ala	TTC Phe	TTC Phe	GAC Asp 520	1590
ACT Thr	GCC Ala	TCA Ser	ACT Thr	GGA Gly 525	AAA Lys	ACA Thr	TTC Phe	CCA Pro	GGT Gly 530	TTC Phe	TTC Phe	TCA Ser	CCT Pro	ATG Met 535	TTA Leu	1638
GGA Gly	GAG Glu	TTT Phe	GTC Val 540	AGT Ser	GAG Glu	ACT Thr	GAG Glu	TCT Ser 545	AGG Arg	GGC Gly	TCA Ser	GAA Glu	TCT Ser 550	GGC Gly	ATC Ile	1686
TTC Phe	ACA Thr	AAT Asn 555	ACA Thr	AAG Lys	GAA Glu	TCC Ser	AGT Ser 560	TCT Ser	CAT His	CAC His	CCT Pro	GGG Gly 565	ATA Ile	GCT Ala	GAA Glu	1734
TTC Phe	CCT Pro 570	TCC Ser	CGT Arg	GGT Gly	AAA Lys	TCT Ser 575	TCA Ser	AGT Ser	TAC Tyr	AGC Ser	AAA Lys 580	CAA Gln	TTT Phe	ACT Thr	AGT Ser	1782
AGC Ser 585	ACG Thr	AGT Ser	TAC Tyr	AAC Asn	AGA Arg 59 0	GGA Gly	GAC Asp	TCC Ser	ACA Thr	TTT Phe 595	GAA Glu	AGC Ser	AAG Lys	AGC Ser	TAT Tyr 600	1830
AAA Lys	ATG Met	GCA Ala	GAT Asp	GAG G1u 605	GCC Ala	GGA Gly	AGT Ser	GAA Glu	GCC Ala 610	GAT Asp	CAT His	GAA Glu	GGA Gly	ACA Thr 615	CAT His	1878
AGC Ser	ACC Thr	AAG Lys	AGA Arg 620	GGG Gly	CAT His	GCT Ala	AAA Lys	TCT Ser 625	CGC Arg	CCT Pro	GTC Val	AGA Arg	GGT Gly 630	ATC Ile	CAC His	1926
ACT Thr	TCT Ser	CCT Pro 635	TTG Leu	GGG Gly	AAG Lys	Pro	TCC Ser 640	CTG Leu	TCC Ser	CCC Pro	TAGA	СТАА	GT T	AAAT	ATTTC	1979
TGCA	CAGT	GT T	CCCA	TGGC	с сс	TTGC	ATTT	ССТ	тстт	AAC	тстс	TGTT	AC A	CGTC	ATTGA	2039
AACT	ACAC	TTT	TTTG	GTCT	G TT	TTTG	TGCT	AGA	CTGT	AAG	TTCC	TTGG	GG G	CAGG	GCCTT	2099
TGTC	TGTC	TC A	тстс	TGTA	T TC	CCAA	ATGC	СТА	ACAG	TAC	AGAG	CCAT	GA C	TCAA	TAAAT	2159
ACAT	GTTA	AA T	GGAT	GAAT	G											2179

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 643 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Phe Ser Met Arg Ile Val Cys Leu Val Leu Ser Val Val Gly Thr 1 5 10 15

Ala Trp Thr Ala Asp Ser Gly Glu Gly Asp Phe Leu Ala Glu Gly Gly
20 25 30

Gly Val Arg Gly Pro Arg Val Val Glu Arg His Gln Ser Ala Cys Lys
35 40 45

Asp Ser Asp Trp Pro Phe Cys Ser Asp Glu Asp Trp Asn Tyr Lys Cys
50
60

Pro Ser Gly Cys Arg Met Lys Gly Leu Ile Asp Glu Val Asn Gln Asp 65 70 75 80

Phe Thr Asn Arg Ile Asn Lys Leu Lys Asn Ser Leu Phe Glu Tyr Gln
85 90 95

Lys Asn Asn Lys Asp Ser His Ser Leu Thr Thr Asn Ile Met Glu Ile 100 105 110

Leu Arg Gly Asp Phe Ser Ser Ala Asn Asn Arg Asp Asn Thr Tyr Asn 115 120 125

Arg Val Ser Glu Asp Leu Arg Ser Arg Ile Glu Val Leu Lys Arg Lys 130 140

Val Ile Glu Lys Val Gln His Ile Gln Leu Leu Gln Lys Asn Val Arg 145 150 155 160

Ala Gln Leu Val Asp Met Lys Arg Leu Glu Val Asp Ile Asp Ile Lys 165 170 175

Ile Arg Ser Cys Arg Gly Ser Trp Ser Arg Ala Leu Ala Arg Glu Val 180 185 190

Asp Leu Lys Asp Tyr Glu Asp Gln Gln Lys Gln Leu Glu Gln Val Ile 195 200 205

Ala Lys Asp Leu Leu Pro Ser Arg Asp Arg Gln His Leu Pro Leu Ile 210 215 220

Lys Met Lys Pro Val Pro Asp Leu Val Pro Gly Asn Phe Lys Ser Gln 225 230 235 240

Leu Gln Lys Val Pro Pro Glu Trp Lys Ala Leu Thr Asp Met Pro Gln Met Arg Met Glu Leu Glu Arg Pro Gly Gly Asn Glu Ile Thr Arg Gly Gly Ser Thr Ser Tyr Gly Thr Gly Ser Glu Thr Glu Ser Pro Arg Asn 280 Pro Ser Ser Ala Gly Ser Trp Asn Ser Gly Ser Ser Gly Pro Gly Ser Thr Gly Asn Arg Asn Pro Gly Ser Ser Gly Thr Gly Gly Thr Ala Thr Trp Lys Pro Gly Ser Ser Gly Pro Gly Ser Ala Gly Ser Trp Asn Ser Gly Ser Ser Gly Thr Gly Ser Thr Gly Asn Gln Asn Pro Gly Ser Pro Arg Pro Gly Ser Thr Gly Thr Trp Asn Pro Gly Ser Ser Glu Arg Gly 360 Ser Ala Gly His Trp Thr Ser Glu Ser Ser Val Ser Gly Ser Thr Gly 375 Gln Trp His Ser Glu Ser Gly Ser Phe Arg Pro Asp Ser Pro Gly Ser 390 Gly Asn Ala Arg Pro Asn Asn Pro Asp Trp Gly Thr Phe Glu Glu Val 410 Ser Gly Asn Val Ser Pro Gly Thr Arg Arg Glu Tyr His Thr Glu Lys Leu Val Thr Lys Gly Asp Lys Glu Leu Arg Thr Gly Lys Glu Lys Val Thr Ser Gly Ser Thr Thr Thr Arg Arg Ser Cys Ser Lys Thr Val 455 Thr Lys Thr Val Ile Gly Pro Asp Gly His Lys Glu Val Thr Lys Glu 470 Val Val Thr Ser Glu Asp Gly Ser Asp Cys Pro Glu Ala Met Asp Leu Gly Thr Leu Ser Gly Ile Gly Thr Leu Asp Gly Phe Arg His Arg His 500 Pro Asp Glu Ala Ala Phe Phe Asp Thr Ala Ser Thr Gly Lys Thr Phe 515 520 525

Pro	Gly 530	Phe	Phe	Ser	Pro	Met 535	Leu	Gly	Glu	Phe	Val 540	Ser	Glu	Thr	G1 c
Ser 545	Arg	Gly	Ser	G1 u	Ser 550	Gly	Ile	Phe	Thr	Asn 555	Thr	Lys	Glu	Ser	Ser 560
Ser	His	His	Pro	Gly 565	Ile	Ala	Glu	Phe	Pro 570	Ser	Arg	Gly	Lys	Ser 575	Ser
Ser	Tyr	Ser	Lys 58 0	Gln	Phe	Thr	Ser	Ser 585	Thr	Ser	Tyr	Asn	Arg 590	Gly	Asp
Ser	Thr	Phe 595	Glu	Ser	Lys	Ser	Tyr 600	Lys	Met	Ala	Asp	Glu 605	Ala	Gly	Ser
Glu	Ala 610	Asp	His	G1 u	Gly	Thr 615	His	Ser	Thr	Lys	Arg 620	Gly	His	Ala	Lys
Ser 625	Arg	Pro	Val	Arg	Gly 630	Пе	His	Thr	Ser	Pro 63 5	Leu	Gly	Lys	Pro	Ser 640
Leu	Ser	Pro													

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 4027 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 3..4013

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

AC ATG GCA GTG AGT CAT GGG AGG GAG AGC AAG CCT CTG ACT GCT C Met Ala Val Ser His Gly Arg Glu Ser Lys Pro Leu Thr Ala G 1 5 10	AA 47 iln 15
CAG ACA ACC AAA CTG GAT GCT CCC ACT AAC CTC CAG TTT GTC AAT Gln Thr Thr Lys Leu Asp Ala Pro Thr Asn Leu Gln Phe Val Asn 20 25 30	GAA 95 Glu
ACT GAT TCT ACT GTC CTG GTG AGA TGG ACT CCA CCT CGG GCC CAG Thr Asp Ser Thr Val Leu Val Arg Trp Thr Pro Pro Arg Ala Gln 35	ATA 143 Ile

191
239
287
335
383
431
479
527
575
623
671
719
767
815

	ACT Thr	GTC Val	AAG Lys	GAT Asp 275	GAC Asp	AAG Lys	GAA Glu	AGT Ser	GTC Val 280	CCT Pro	ATC Ile	TCT Ser	GAT Asp	ACC Thr 285	ATC Ile	ATC Ile	863
						CTC Leu											911
	TCA Ser	AGC Ser 305	ATC Ile	GGC Gly	CTG Leu	AGG Arg	TGG Trp 310	ACC Thr	CCG Pro	CTA Leu	AAC Asn	TCT Ser 315	TCC Ser	ACC Thr	ATT Ile	ATT Ile	959
						GTA Val 325											1007
	GAA Glu	GAT Asp	TTT Phe	GTG Val	TAC Tyr 340	TCC Ser	TCA Ser	GTA Val	GGA Gly	TAC Tyr 345	TAC Tyr	ACA Thr	GTC Val	ACA Thr	GGG Gly 350	CTG Leu	1055
						TAT Tyr											1103
						ACT Thr											1151
						TTC Phe											1199
	ACC Thr 400	TGG Trp	GCT Ala	CCA Pro	CCC Pro	CCA Pro 405	TCC Ser	ATT Ile	GAT Asp	TTA Leu	ACC Thr 410	AAC Asn	TTC Phe	CTG Leu	GTG Val	CGT Arg 415	1247
•						AAT Asn											1295
						GTG Val											1343
						TCC Ser											1391
	CTT Leu	AGA Arg 4 65	GGA Gly	AGA Arg	CAG Gln	AAA Lys	ACA Thr 470	GGT Gly	CTT Leu	GAT Asp	TCC Ser	CCA Pro 475	ACT Thr	GGC Gly	ATT Ile	GAC Asp	1439

TT1 Phe 480	3 2 6 I	GAT Asp	T ATT	ACT Thr	GCC Ala 485	ı Asr	C TCT Ser	TTT Phe	ACT Thr	GTG Val 490	His	TG(Trp	ATT Ile	GCT Ala	CCT Pro 495	1487	7
CG <i>A</i> Arg	A GCC Ala	ACC Thr	TTG TTG	ACT Thr 500	Gly	TAC Tyr	AGG Arg	ATC Ile	CGC Arg 505	His	CAT	CCC Pro	GAC Glu	G CAC His 510	TTC Phe	1535	5
AGT Ser	GGG Gly	AGA Arg	CCT Pro 515	Arg	GAA Glu	GAT Asp	CGG Arg	GTG Val 520	Pro	CAC His	TCT Ser	CGG Arg	AA1 Asr 525	Ser	ATC	1583	3
ACC Thr	CTC Leu	ACC Thr 530	' Asn	CTC Leu	ACT Thr	CCA Pro	GGC Gly 535	Thr	GAG Glu	TAT Tyr	GTG Val	GTC Val 540	Ser	ATC	GTT Val	1631	
GCT Ala	CTT Leu 545	Asn	GGC Gly	AGA Arg	GAG Glu	GAA G1u 550	Ser	CCC Pro	TTA Leu	TTG Leu	ATT Ile 555	Gly	CAA Gln	CAA Gln	TCA Ser	1679	ı
ACA Thr 560	vai	TCT Ser	GAT Asp	GTT Val	CCG Pro 565	Arg	GAC Asp	CTG Leu	GAA Glu	GTT Val 570	GTT Val	GCT Ala	GCG Ala	ACC Thr	CCC Pro 575	1727	
ACC Thr	AGC Ser	CTA Leu	CTG Leu	ATC Ile 580	AGC Ser	TGG Trp	GAT Asp	GCT Ala	CCT Pro 585	GCT Ala	GTC Val	ACA Thr	GTG Val	AGA Arg 590	TAT Tyr	1775	
TAC Tyr	AGG Arg	ATC Ile	ACT Thr 595	TAC Tyr	GGA Gly	GAA Glu	ACA Thr	GGA Gly 600	GGA Gly	AAT Asn	AGC Ser	CCT Pro	GTC Val 605	CAG Gln	GAG Glu	1823	
TTC Phe	ACT Thr	GTG Val 610	CCT Pro	GGG Gly	AGC Ser	AAG Lys	TCT Ser 615	ACA Thr	GCT Ala	ACC Thr	ATC Ile	AGC Ser 620	GGC Gly	CTT Leu	AAA Lys	1871	
CCT Pro	GGA Gly 625	GTT Val	GAT Asp	TAT Tyr	ACC Thr	ATC Ile 630	ACT Thr	GTG Val	TAT Tyr	GCT Ala	GTC Val 635	ACT Thr	GGC Gly	CGT Arg	GGA Gly	1919	
GAC Asp 640	AGC Ser	CCC Pro	GCA Ala	AGC Ser	AGC Ser 645	AAG Lys	CCA Pro	ATT Ile	TCC Ser	ATT Ile 650	AAT Asn	TAC Tyr	CGA Arg	ACA Thr	GAA Glu 655	1967	
ATT Ile	GAC Asp	AAA Lys	CCA Pro	TCC Ser 660	CAG Gln	ATG Met	CAA Gln	GTG Val	ACC Thr 665	GAT Asp	GTT Val	CAG Gln	GAC Asp	AAC Asn 670	AGC Ser	2015	
ATT Ile	AGT Ser	GTC Val	AAG Lys 675	TGG Trp	CTG Leu	CCT Pro	TCA Ser	AGT Ser 680	TCC Ser	CCT Pro	GTT Val	ACT Thr	GGT Gly 685	TAC Tyr	AGA Arg	2063	
GTA Val	inr	ACC Thr 690	ACT Thr	CCC Pro	AAA Lys	Asn	GGA Gly 695	CCA Pro	GGA Gly	CCA Pro	ACA Thr	AAA Lys 700	ACT Thr	AAA Lys	ACT Thr	2111	

GCA Ala	GGT Gly 705	Pro	GAT Asp	CAA Gln	ACA Thr	GAA Glu 710	Met	ACT Thr	ATT Ile	GAA Glu	GGC Gly 715	Leu	CAG Gln	CCC Pro	ACA Thr		2159
GTG Val 720	GAG Glu	TAT Tyr	GTG Val	GTT Val	AGT Ser 725	GTC Val	TAT Tyr	GCT Ala	CAG Gln	AAT Asn 730	CCA Pro	AGC Ser	GGA Gly	GAG Glu	AGT Ser 735		2207
CAG Gln	CCT Pro	CTG Leu	GTT Val	CAG Gln 740	ACT Thr	GCA Ala	GTA Val	ACC Thr	AAC Asn 745	ATT Ile	GAT Asp	CGC Arg	CCT Pro	AAA Lys 750			2255
CTG Leu	GCA Ala	TTC Phe	ACT Thr 755	GAT Asp	GTG Val	GAT Asp	GTC Val	GAT Asp 760	TCC Ser	ATC Ile	AAA Lys	ATT Ile	GCT Ala 765	TGG Trp	GAA Glu		2303
AGC Ser	CCA Pro	CAG Gln 770	GGG Gly	CAA Gln	GTT Val	TCC Ser	AGG Arg 775	TAC Tyr	AGG Arg	GTG Val	ACC Thr	TAC Tyr 780	TCG Ser	AGC Ser	CCT Pro		2351
GAG Glu	GAT Asp 785	GGA Gly	ATC Ile	CAT His	GAG G1u	CTA Leu 790	TTC Phe	CCT Pro	GCA Ala	CCT Pro	GAT Asp 795	GGT Gly	GAA Glu	GAA Glu	GAC Asp		2399
ACT Thr 800	GCA Ala	GAG Glu	CTG Leu	CAA Gln	GGC Gly 805	CTC Leu	AGA Arg	CCG Pro	GGT Gly	TCT Ser 810	GAG Glu	TAC Tyr	ACA Thr	GTC Val	AGT Ser 815	٠.	2447
GTG Val	GTT Val	GCC Ala	TTG Leu	CAC His 820	GAT Asp	GAT Asp	ATG Met	GAG G1u	AGC Ser 825	CAG Gln	CCC Pro	CTG Leu	ATT Ile	GGA Gly 830	ACC Thr		2495
CAG Gln	TCC Ser	ACA Thr	GCT Ala 835	ATT Ile	CCT Pro	GCA Ala	CCA Pro	ACT Thr 840	GAC Asp	CTG Leu	AAG Lys	TTC Phe	ACT Thr 845	CAG Gln	GTC Val		2543
ACA Thr	CCC Pro	ACA Thr 850	AGC Ser	CTG Leu	AGC Ser	GCC Ala	CAG G1n 855	TGG Trp	ACA Thr	CCA Pro	CCC Pro	AAT Asn 860	GTT Val	CAG Gln	CTC Leu		2591
inr	GGA Gly 865	TAT Tyr	CGA Arg	GTG Val	CGG Arg	GTG Val 870	ACC Thr	CCC Pro	AAG Lys	GAG Glu	AAG Lys 875	ACC Thr	GGA Gly	CCA Pro	ATG Met		2639
AAA Lys 880	GAA Glu	ATC Ile	AAC Asn	CTT Leu	GCT Ala 885	CCT Pro	GAC Asp	AGC Ser	TCA Ser	TCC Ser 890	GTG Val	GTT Val	GTA Val	TCA Ser	GGA Gly 895		2687
CTT Leu	ATG Met	GTG Val	GCC Ala	ACC Thr 900	AAA Lys	TAT Tyr	GAA Glu	GTG Val	AGT Ser 905	GTC Val	TAT Tyr	GCT Ala	CTT Leu	AAG Lys 910	GAC Asp		2735

ACT Thr	TTG Leu	ACA Thr	AGC Ser 915	AGA Arg	CCA Pro	GCT Ala	CAG Gln	GGT Gly 920	GTT Val	GTC Val	ACC Thr	ACT Thr	CTG Leu 925	GAG Glu	GGA Gly	2783
GGA Gly	AAT Asn	TTT Phe 930	Lys	AGC Ser	CAG G1n	CTT Leu	CAG Gln 935	AAG Lys	GTA Val	CCC Pro	CCA Pro	GAG Glu 940	TGG Trp	AAG Lys	GCA Ala	2831
TTA Leu	ACA Thr 945	Asp	ATG Met	CCG Pro	CAG Gln	ATG Met 950	AGA Arg	ATG Met	GAG Glu	TTA Leu	GAG G1u 95 5	AGA Arg	CCT Pro	GGT Gly	GGA Gly	2879
AAT Asn 960	Glu	ATT Ile	ACT Thr	CGA Arg	GGA Gly 965	GGC Gly	TCC Ser	ACC Thr	TCT Ser	TAT Tyr 970	GGA Gly	ACC Thr	GGA Gly	TCA Ser	GAG Glu 975	2927
ACG Thr	GAA Glu	AGC Ser	CCC Pro	AGG Arg 980	AAC Asn	CCT Pro	AGC Ser	AGT Ser	GCT Ala 985	GGA Gly	AGC Ser	TGG Trp	AAC Asn	TCT Ser 990	GGG Gly	2975
AGC Ser	TCT Ser	GGA Gly	CCT Pro 995	GGA Gly	AGT Ser	ACT Thr	GGA Gly	AAC Asn 1000	Arg	AAC Asn	CCT Pro	GGG Gly	AGC Ser 100	Ser	GGG Gly	3023
ACT Thr	GGA Gly	GGG Gly 1010	Thr	GCA Ala	ACC Thr	TGG Trp	AAA Lys 1015	Pro	GGG Gly	AGC Ser	TCT Ser	GGA Gly 1020	Pro	GGA Gly	AGT Ser	3071
GCT Ala	GGA Gly 102	Ser	TGG Trp	AAC Asn	TCT Ser	GGG Gly 1030	Ser	TCT Ser	GGA Gly	ACT Thr	GGA Gly 103!	AGT Ser	ACT Thr	GGA Gly	AAC Asn	3119
CAA Gln 1040	Asn	CCT Pro	GGG Gly	AGC Ser	CCT Pro 1045	Arg	CCT Pro	GGT Gly	AGT Ser	ACC Thr 1050	Gly	ACC Thr	TGG Trp	AAT Asn	CCT Pro 1055	3167
GGC Gly	AGC Ser	TCT Ser	GAA Glu	CGC Arg 1060	Gly	AGT Ser	GCT Ala	GGG Gly	CAC His 1065	Trp	ACC Thr	TCT Ser	GAG Glu	AGC Ser 1070	Ser	3215
GTA Val	TCT Ser	GGT Gly	AGT Ser 1075	Ihr	GGA Gly	CAA Gln	TGG Trp	CAC His 1080	Ser	GAA Glu	TCT Ser	GGA Gly	AGT Ser 1085	Phe	AGG Arg	3263
CCA Pro	GAT Asp	AGC Ser 1090	Pro	GGC Gly	TCT Ser	GGG Gly	AAC Asn 1095	Ala	AGG Arg	CCT Pro	AAC Asn	AAC Asn 1100	Pro	GAC Asp	TGG Trp	3311
GGC Gly	ACA Thr 1105	Phe	GAA Glu	GAG Glu	GTG Val	TCA Ser 1110	Gly	AAT Asn	GTA Val	AGT Ser	CCA Pro 1115	GGG Gly	ACA Thr	AGG Arg	AGA Arg	3359
GAG Glu 1120	Tyr	CAC His	ACA Thr	GAA Glu	AAA Lys 1125	Leu	GTC Val	ACT Thr	AAA Lys	GGA Gly 1130	Asp	AAA Lys	GAG Glu	CTC Leu	AGG Arg 1135	3407

ACT GGT AAA GAG Thr Gly Lys Glu			Thr Thr Thr	
TCA TGC TCT AAA Ser Cys Ser Lys 1155	Thr Val Thr Ly	AG ACT GTT ATT ys Thr Val Ile 1160	GGT CCT GAT (Gly Pro Asp (1165	GGT CAC 3503 Gly His
AAA GAA GTT ACC Lys Glu Val Thr 1170	Lys Glu Val Va			
CCC GAG GCA ATG Pro Glu Ala Met 1185				
GGG TTC CGC CAT Gly Phe Arg His 1200			Phe Phe Asp	
TCA ACT GGA AAA Ser Thr Gly Lys			Pro Met Leu (
TTT GTC AGT GAG Phe Val Ser Glu 1235	Thr Glu Ser Ar			
AAT ACA AAG GAA Asn Thr Lys Glu 1250	Ser Ser Ser Hi			
TCC CGT GGT AAA Ser Arg Gly Lys 1265		r Ser Lys Gln		
AGT TAC AAC AGA Ser Tyr Asn Arg 1280			Lys Ser Tyr I	
GCA GAT GAG GCC Ala Asp Glu Ala	GGA AGT GAA GO Gly Ser Glu Al 1300	CC GAT CAT GAA la Asp His Glu 1305	Gly Thr His !	AGC ACC 3935 Ser Thr 1310
AAG AGA GGC CAT Lys Arg Gly His 1315	Ala Lys Ser Ar			
CCT TTG GGG AAG Pro Leu Gly Lys 1330	Pro Ser Leu Se		GT TAAATAT	4027

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1336 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:
- Met Ala Val Ser His Gly Arg Glu Ser Lys Pro Leu Thr Ala Gln Gln
 1 10 15
- Thr Thr Lys Leu Asp Ala Pro Thr Asn Leu Gln Phe Val Asn Glu Thr 20 25 30
- Asp Ser Thr Val Leu Val Arg Trp Thr Pro Pro Arg Ala Gln Ile Thr 35 40 45
- Gly Tyr Arg Leu Thr Val Gly Leu Thr Arg Arg Gly Gln Pro Arg Gln
 50 60
- Tyr Asn Val Gly Pro Ser Val Ser Lys Tyr Pro Leu Arg Asn Leu Gln
 65 70 75 80
- Pro Ala Ser Glu Tyr Thr Val Ser Leu Val Ala Ile Lys Gly Asn Gln
 85 90 95
- Glu Ser Pro Lys Ala Thr Gly Val Phe Thr Thr Leu Gln Pro Gly Ser
- Ser Ile Pro Pro Tyr Asn Thr Glu Val Thr Glu Thr Thr Ile Val Ile 115 120 125
- Thr Trp Thr Pro Ala Pro Arg Ile Gly Phe Lys Leu Gly Val Arg Pro 130 135 140
- Ser Gln Gly Glu Ala Pro Arg Glu Val Thr Ser Asp Ser Gly Ser 145 150 155 160
- Ile Val Val Ser Gly Leu Thr Pro Gly Val Glu Tyr Val Tyr Thr Ile
 165 170 175
- Gln Val Leu Arg Asp Gly Gln Glu Arg Asp Ala Pro Ile Val Asn Lys 180 185 190
- Val Val Thr Pro Leu Ser Pro Pro Thr Asn Leu His Leu Glu Ala Asn 195 200 205
- Pro Asp Thr Gly Val Leu Thr Val Ser Trp Glu Arg Ser Thr Thr Pro 210 215 220
- Asp Ile Thr Gly Tyr Arg Ile Thr Thr Thr Pro Thr Asn Gly Gln Gln 225 230 235 240

Gly Asn Ser Leu Glu Glu Val Val His Ala Asp Gln Ser Ser Cys Thr Phe Asp Asn Leu Ser Pro Gly Leu Glu Tyr Asn Val Ser Val Tyr Thr 265 Val Lys Asp Asp Lys Glu Ser Val Pro Ile Ser Asp Thr Ile Ile Pro Glu Val Pro Gln Leu Thr Asp Leu Ser Phe Val Asp Ile Thr Asp Ser 290 Ser Ile Gly Leu Arg Trp Thr Pro Leu Asn Ser Ser Thr Ile Ile Gly 315 Tyr Arg Ile Thr Val Val Ala Ala Gly Glu Gly Ile Pro Ile Phe Glu 335 Asp Phe Val Tyr Ser Ser Val Gly Tyr Tyr Thr Val Thr Gly Leu Glu 345 Pro Gly Ile Asp Tyr Asp Ile Ser Val Ile Thr Leu Ile Asn Gly Gly 360 Glu Ser Ala Pro Thr Thr Leu Thr Gln Gln Thr Ala Val Pro Pro Pro 375 Thr Asp Leu Arg Phe Thr Asn Ile Gly Pro Asp Thr Met Arg Val Thr 395 Trp Ala Pro Pro Pro Ser Ile Asp Leu Thr Asn Phe Leu Val Arg Tyr Ser Pro Val Lys Asn Glu Glu Asp Val Ala Glu Leu Ser Ile Ser Pro 425 Ser Asp Asn Ala Val Val Leu Thr Asn Leu Leu Pro Gly Thr Glu Tyr 440 Val Val Ser Val Ser Ser Val Tyr Glu Gln His Glu Ser Thr Pro Leu 455 Arg Gly Arg Gln Lys Thr Gly Leu Asp Ser Pro Thr Gly Ile Asp Phe 475 Ser Asp Ile Thr Ala Asn Ser Phe Thr Val His Trp Ile Ala Pro Arg 485 490 Ala Thr Ile Thr Gly Tyr Arg Ile Arg His His Pro Glu His Phe Ser 500 505 Gly Arg Pro Arg Glu Asp Arg Val Pro His Ser Arg Asn Ser Ile Thr

520

525

Leu Thr Asn Leu Thr Pro Gly Thr Glu Tyr Val Val Ser Ile Val Ala 530 540

Leu Asn Gly Arg Glu Glu Ser Pro Leu Leu Ile Gly Gln Gln Ser Thr 545 550 550

Val Ser Asp Val Pro Arg Asp Leu Glu Val Val Ala Ala Thr Pro Thr 565 570 575

Ser Leu Leu Ile Ser Trp Asp Ala Pro Ala Val Thr Val Arg Tyr Tyr 580 585 590

Arg Ile Thr Tyr Gly Glu Thr Gly Gly Asn Ser Pro Val Gln Glu Phe 595 600 605

Thr Val Pro Gly Ser Lys Ser Thr Ala Thr Ile Ser Gly Leu Lys Pro 610 620

Gly Val Asp Tyr Thr Ile Thr Val Tyr Ala Val Thr Gly Arg Gly Asp 625 635 640

Ser Pro Ala Ser Ser Lys Pro Ile Ser Ile Asn Tyr Arg Thr Glu Ile 645 650 655

Asp Lys Pro Ser Gln Met Gln Val Thr Asp Val Gln Asp Asn Ser Ile 660 665 670

Ser Val Lys Trp Leu Pro Ser Ser Ser Pro Val Thr Gly Tyr Arg Val 675 680 685

Thr Thr Pro Lys Asn Gly Pro Gly Pro Thr Lys Thr Lys Thr Ala
690 695 700

Gly Pro Asp Gln Thr Glu Met Thr Ile Glu Gly Leu Gln Pro Thr Val 705 710 715 720

Glu Tyr Val Val Ser Val Tyr Ala Gln Asn Pro Ser Gly Glu Ser Gln 725 730 735

Pro Leu Val Gln Thr Ala Val Thr Asn Ile Asp Arg Pro Lys Gly Leu
740 745 750

Ala Phe Thr Asp Val Asp Val Asp Ser Ile Lys Ile Ala Trp Glu Ser 755 760 765

Pro Gln Gly Gln Val Ser Arg Tyr Arg Val Thr Tyr Ser Ser Pro Glu 770 780

Asp Gly Ile His Glu Leu Phe Pro Ala Pro Asp Gly Glu Glu Asp Thr 785 790 795 800

Ala Glu Leu Gln Gly Leu Arg Pro Gly Ser Glu Tyr Thr Val Ser Val 805 810 815 Val Ala Leu His Asp Asp Met Glu Ser Gln Pro Leu Ile Gly Thr Gln 820 825 830

Ser Thr Ala Ile Pro Ala Pro Thr Asp Leu Lys Phe Thr Gln Val Thr 835 840 845

Pro Thr Ser Leu Ser Ala Gln Trp Thr Pro Pro Asn Val Gln Leu Thr 850 855 860

Gly Tyr Arg Val Arg Val Thr Pro Lys Glu Lys Thr Gly Pro Met Lys 865 870 875 880

Glu Ile Asn Leu Ala Pro Asp Ser Ser Ser Val Val Ser Gly Leu 885 890 895

Met Val Ala Thr Lys Tyr Glu Val Ser Val Tyr Ala Leu Lys Asp Thr 900 905 910

Leu Thr Ser Arg Pro Ala Gln Gly Val Val Thr Thr Leu Glu Gly Gly 915 920 925

Asn Phe Lys Ser Gln Leu Gln Lys Val Pro Pro Glu Trp Lys Ala Leu 930 - 935 940

Thr Asp Met Pro Gln Met Arg Met Glu Leu Glu Arg Pro Gly Gly Asn 945 950 955 960

Glu Ile Thr Arg Gly Gly Ser Thr Ser Tyr Gly Thr Gly Ser Glu Thr 965 970 975

Glu Ser Pro Arg Asn Pro Ser Ser Ala Gly Ser Trp Asn Ser Gly Ser 980 985 990

Ser Gly Pro Gly Ser Thr Gly Asn Arg Asn Pro Gly Ser Ser Gly Thr 995 1000 1005

Gly Gly Thr Ala Thr Trp Lys Pro Gly Ser Ser Gly Pro Gly Ser Ala 1010 1015 1020

Gly Ser Trp Asn Ser Gly Ser Ser Gly Thr Gly Ser Thr Gly Asn Gln 1025 1030 1035 1040

Asn Pro Gly Ser Pro Arg Pro Gly Ser Thr Gly Thr Trp Asn Pro Gly 1045 1050 1055

Ser Ser Glu Arg Gly Ser Ala Gly His Trp Thr Ser Glu Ser Ser Val 1060 1065 1070

Ser Gly Ser Thr Gly Gln Trp His Ser Glu Ser Gly Ser Phe Arg Pro 1075 1080 1085

Asp Ser Pro Gly Ser Gly Asn Ala Arg Pro Asn Asn Pro Asp Trp Gly 1090 1095 1100

Thr Phe Glu Glu Val Ser Gly Asn Val Ser Pro Gly Thr Arg Arg Glu 1105 1110 1115 1120

Tyr His Thr Glu Lys Leu Val Thr Lys Gly Asp Lys Glu Leu Arg Thr 1125 1130 1135

Gly Lys Glu Lys Val Thr Ser Gly Ser Thr Thr Thr Arg Arg Ser 1140 1145 1150

Cys Ser Lys Thr Val Thr Lys Thr Val Ile Gly Pro Asp Gly His Lys 1155 1160 1165

Glu Val Thr Lys Glu Val Val Thr Ser Glu Asp Gly Ser Asp Cys Pro 1170 1175 1180

Glu Ala Met Asp Leu Gly Thr Leu Ser Gly Ile Gly Thr Leu Asp Gly 1185 1200

Phe Arg His Arg His Pro Asp Glu Ala Ala Phe Phe Asp Thr Ala Ser 1205 1210 1215

Thr Gly Lys Thr Phe Pro Gly Phe Phe Ser Pro Met Leu Gly Glu Phe 1220 1235 1230

Val Ser Glu Thr Glu Ser Arg Gly Ser Glu Ser Gly Ile Phe Thr Asn 1235 1240 1245

Thr Lys Glu Ser Ser His His Pro Gly Ile Ala Glu Phe Pro Ser 1250 1255 1260

Arg Gly Lys Ser Ser Ser Tyr Ser Lys Gln Phe Thr Ser Ser Thr Ser 1265 1270 1275 1280

Tyr Asn Arg Gly Asp Ser Thr Phe Glu Ser Lys Ser Tyr Lys Met Ala 1285 1290 1295

Asp Glu Ala Gly Ser Glu Ala Asp His Glu Gly Thr His Ser Thr Lys 1300 1305 1310

Arg Gly His Ala Lys Ser Arg Pro Val Arg Gly Ile His Thr Ser Pro 1315 1320 1325

Leu Gly Lys Pro Ser Leu Ser Pro 1330 1335

(2) INFOR	RMATION FOR SEQ ID NO:7:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 27 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(vii)	IMMEDIATE SOURCE: (B) CLONE: ZC1551	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:7:	
GATCCCCGG	G GAGCTCCTCG AGGCATG	27
(2) INFOR	MATION FOR SEQ ID NO:8:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 19 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(vii)	IMMEDIATE SOURCE: (B) CLONE: ZC1552	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:8:	
CCTCGAGGA	G CTCCCCGGG	19
(2) INFOR	MATION FOR SEQ ID NO:9:	
(i) :	SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(vii)	IMMEDIATE SOURCE: (B) CLONE: ZC2052	
(xi) \$	SEQUENCE DESCRIPTION: SEQ ID NO:9:	
AATTCACCAT	T GGCAGTGAGT	20

(2) INFORMATION FOR SEQ ID NO:10:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC2053	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:	0:
CATGACTCAC TGCCATGGTG	20
(2) INFORMATION FOR SEQ ID NO:11:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC2491	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1	1:
CTAGATTAGA ATGGGGCC	18
(2) INFORMATION FOR SEQ ID NO:12:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 10 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC2493	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1	2:
CCATTCTAAT	10

. 77

(2) INFORMATION FOR SEQ ID NO:13:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 88 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC3521	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:	
TCGACTTAAG GACACTTTGA CAAGCAGACC AGCTCAGGGT GTTGTCACCA CTCTGGAGGG	60
AGGAAATTTT AAGAGCCAGC TTCAGAAG	88
(2) INFORMATION FOR SEQ ID NO:14:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 88 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC3522	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:	
GTACCTTCTG AAGCTGGCTC TTAAAATTTC CTCCCTCCAG AGTGGTGACA ACACCCTGAG	60
CTGGTCTGCT TGTCAAAGTG TCCTTAAG	88

I Claim:

- 1. A hybrid protein comprising a tissue-binding domain from a first protein covalently linked to a cross-linking domain from a second protein.
- 2. A hybrid protein according to claim 1 wherein the tissue-binding domain of the first protein is a heparin binding domain of thrombospondin, a heparin binding domain of fibronectin, a collagen binding domain of fibronectin or a cell binding domain of fibronectin.
- 3. A hybrid protein according to claim 1 wherein the tissue-binding domain of the first protein comprises the amino acid sequence of Sequence ID No. 6 from Alanine, amino acid 2 to Glutamic acid, amino acid number 926.
- 4. A hybrid protein according to claim 1 wherein the cross-linking domain of the second protein comprises the carboxy-terminal 103 amino acids of loricrin; the ten amino acid repeat beginning with glutamine, amino acid number 496 of involucrin; or the 400 amino-terminal amino acids of the fibrinogen α chain.
- 5. A hybrid protein according to claim 1 wherein the cross-linking domain of the second protein comprises the amino acid sequence of Sequence ID No. 6 from Glycine, amino acid number 928 to Proline, amino acid number 1336.
- 6. A hybrid protein according to claim 1 comprising the amino acid sequence of Sequence ID Number 6 from alanine, amino acid number 2 to Proline, amino acid number 1336.
- 7. An isolated DNA molecule encoding a hybrid protein comprising a first DNA segment encoding a tissue-binding domain from a first protein joined to a second DNA segment encoding a cross-linking domain from a second protein.

- 8. A DNA molecule according to claim 7 wherein the first DNA segment encodes a heparin binding domain of thrombospondin, a heparin binding domain of fibronectin, a collagen binding domain of fibronectin, a collagen binding domain of fibronectin.
- 9. A DNA molecule according to claim 7 wherein the first DNA segment comprises the nucleotide sequence of Sequence ID No. 5 from nucleotide 3 to nucleotide 2780.
- 10. A DNA molecule according to claim 7 wherein the first DNA segment encodes the amino acid sequence of Sequence ID No. 6 from methionine, amino acid number 1 to glutamic acid, amino acid number 926.
- 11. A DNA molecule according to claim 7 wherein the second DNA segment encodes the carboxy-terminal 103 amino acids of loricrin; the ten amino acid repeat beginning with glutamine, amino acid number 496 of involucrin; or the 400 amino-terminal amino acids of the fibrinogen α chain.
- 12. A DNA molecule according to claim 7 wherein the second DNA segment comprises the nucleotide sequence of Sequence ID No. 5 from nucleotide 2784 to nucleotide 4013.
- 13. A DNA molecule according to claim 7 wherein the second DNA segment encodes the amino acid sequence of Sequence ID No. 6 from glycine, amino acid number 928 to proline, amino acid number 1336.
- 14. A DNA molecule according to claim 7 wherein the DNA molecule encodes the amino acid sequence of Sequence ID Number 6 from Methionine, amino acid number 1 to Proline, amino acid number 1336.

- 15. A DNA molecule according to claim 7 wherein the DNA molecule comprises the nucleotide sequence of Sequence ID Number 5 from nucleotide 3 to nucleotide 4013.
- 16. A DNA construct comprising a DNA molecule encoding a hybrid protein, wherein said DNA molecule comprises a first DNA segment encoding a tissue-binding domain from a first protein joined to a second DNA segment encoding a crosslinking domain from a second protein, and wherein said DNA molecule is operably linked to other DNA segments required for the expression of the DNA molecule.
- 17. A DNA construct according to claim 16 wherein the first DNA segment encodes a heparin binding domain of thrombospondin, a heparin binding domain of fibronectin, a collagen binding domain of fibronectin or a cell binding domain of fibronectin.
- 18. A DNA construct according to claim 16 wherein the first DNA segment comprises the nucleotide sequence of Sequence ID No. 5 from nucleotide 3 to nucleotide 2780.
- 19. A DNA construct according to claim 16 wherein the first DNA segment encodes the amino acid sequence of Sequence ID No. 6 from methionine, amino acid 1 to Glutamic acid, amino acid number 926.
- 20. A DNA construct according to claim 16 wherein the second DNA segment encodes the carboxy-terminal 103 amino acids of loricrin; the ten amino acid repeat beginning with glutamine, amino acid number 496 of involucrin; or the 400 amino-terminal amino acids of the fibrinogen α chain.
- 21. A DNA construct according to claim 16 wherein the second DNA segment comprises the nucleotide sequence of Sequence ID No. 5 from nucleotide 2784 to nucleotide 4013.

- 22. A DNA construct according to claim 16 wherein the second DNA segment encodes the amino acid sequence of Sequence ID No. 6 from glycine, amino acid number 928 to proline, amino acid number 1336.
- 23. A DNA construct according to claim 16 wherein the DNA molecule comprises the nucleotide sequence of Sequence ID Number 5 from nucleotide 1 to nucleotide 4013.
- 24. A DNA construct according to claim 16 wherein the DNA molecule encodes the amino acid sequence of Sequence ID Number 6 from Methionine, amino acid number 1 to Proline, amino acid number 1336.
- 25. A host cell containing a DNA construct according to claim 16.
- 26. A method for producing a hybrid protein comprising culturing a host cell according to claim 25 under conditions promoting the expression of the first DNA segment.

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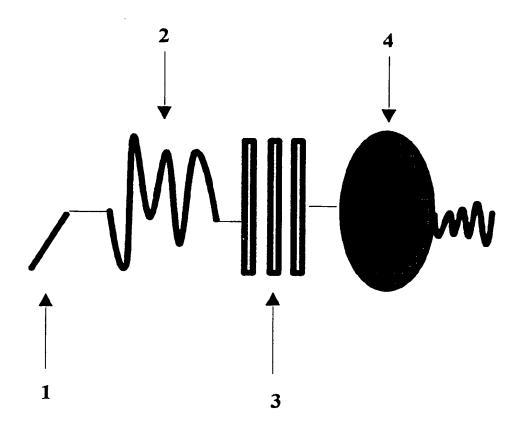


FIGURE 1

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Hybrid+FXIII+Thrombin

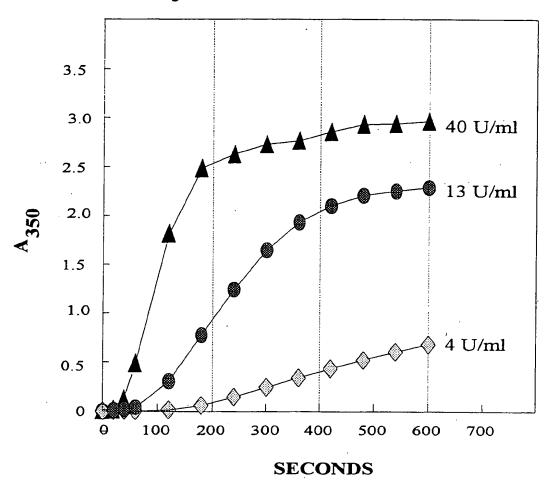


FIGURE 2

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Hybrid+FXIIIa

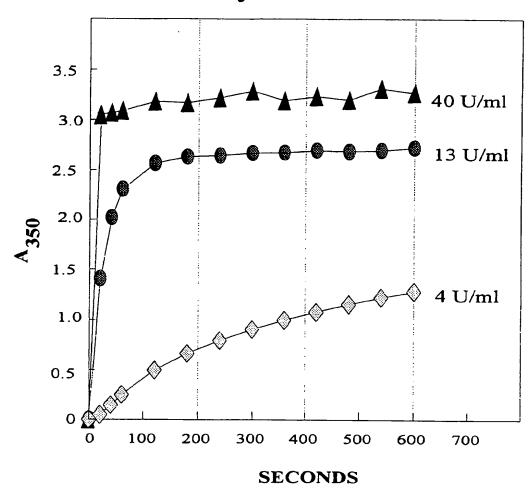


FIGURE 3

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FXIII+Thrombin

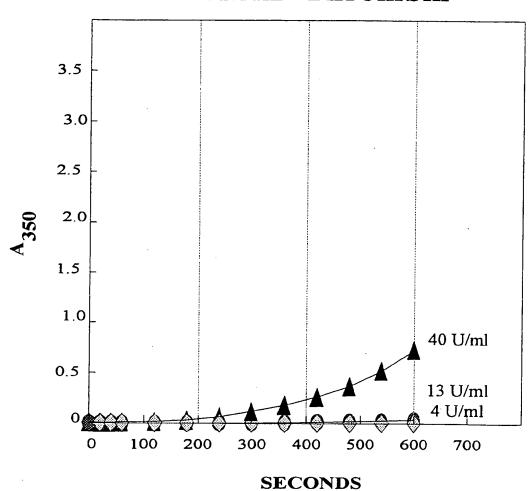


FIGURE 4

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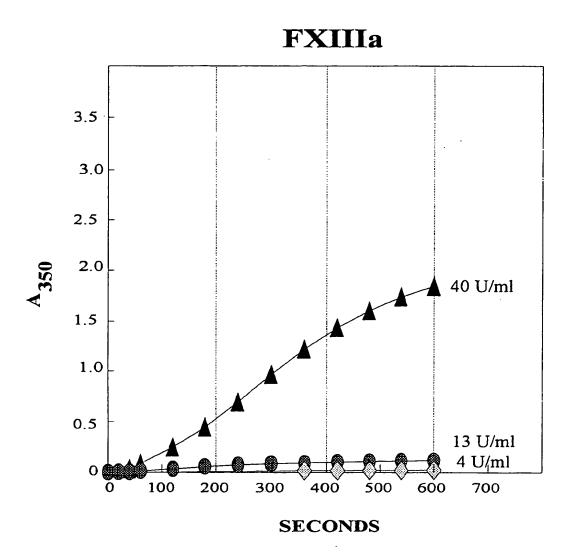


FIGURE 5